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## Antioxidant activity of olive oil mill wastewater obtained from different thermal treatments

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### RESUMEN

#### Actividad antioxidante del alpechín obtenido con diferentes tratamientos térmicos.

En la industria alimentaria, el alpechín se considera un subproducto debido a la presencia de compuestos bioestáticos, con una alta tasa de contaminación, particularmente los fenoles. Además, durante el procesado de la aceituna, la generación de una gran cantidad de este subproducto supone un problema ecológico y económico para los productores. Es importante la reutilización de este agua de desecho para obtener compuestos útiles. Para purificar el agua de desecho es necesario el desarrollo de operaciones que modifiquen su contenido orgánico, para poder obtener agentes fertilizantes y/o recuperar sustancias con un alto valor añadido como los compuestos fenólicos, que actualmente están reconocidos científicamente como moléculas con una elevada actividad antioxidante. Se realizó un análisis cromatográfico de dichos compuestos para caracterizar las diferentes concentraciones en el alpechín, y posteriormente se determinó el poder reductor de los extractos. El tratamiento térmico del alpechín en rotavapor y en estufa incrementó la eficiencia en la captura de radicales. Estos resultados pueden correlacionarse con la posibilidad de recuperar y reutilizar este tipo de desecho debido a sus propiedades antioxidantes.

**PALABRAS CLAVE:** Alpechín – Capacidad antioxidante – Polifenoles – Radical DPPH’.

### SUMMARY

#### Antioxidant activity of olive oil mill wastewater obtained from different thermal treatments.

In food industry, Olive Oil Mill Wastewater (OOMWW) is considered a by-product because of the presence of biostatic compounds with a high polluting rate, in particular phenols. Moreover, during olive oil processing, a large amount of this by-product constitutes an ecological and economical problem for the producers. To reevaluate this by-product, the reuse of this wastewater to obtain useful compounds appears to be very important. In order to purify the wastewater, the development of operations that modify its organic content seems necessary for obtaining of eventual fertilizing agents and/or to recover substances with a high added value such as phenolic compounds, which are currently recognized scientifically as molecules with a high antioxidant activity. A chromatographic analysis of these compounds was conducted to characterize different concentrations of wastewater and the reducing power of the extracts was measured. The thermal treatment of olive oil mill wastewater in a rotary evaporator and in an oven involved an increase in

radical scavenging efficiency. These results could be correlated with the possibility of recovering and reusing this type of waste for its antioxidant properties.

**KEY-WORDS:** Antioxidant capacity – DPPH’ radical – Olive oil mill waste water – Polyphenols.

### 1. INTRODUCTION

Olives and olive oil are foods which are particularly rich in antioxidant biomolecules, named polyphenols. The olive oil market has recently expanded, since the “Mediterranean diet” is widely appreciated throughout the World by consumers who pay more attention to both health and nutritional food aspects (Key, 1995).

Olive processing produces OOMWW, which represents the main environmental problem and consists of several vegetable compounds (mineral but particularly organic substances), coming from the olive pulp through olive oil processing and equipment wash water and diluting water for olive paste in a continuous extraction system (three-phases decanter). Continuous three-phase extraction systems are still widely used in olive oil mills, especially in Italy, where in most cases they have not yet been replaced by more recent two-phase extraction systems, which involve a reduced volume of OOMWW but an increased concentration in organic matter (Roig *et al.*, 2006). The three-phase extraction system involves the addition of large amounts of water (up to 50 L 100 Kg<sup>-1</sup> olive paste) resulting in the worldwide production of more than 30 million m<sup>3</sup> per year of OOMWW (Borja *et al.*, 1997). Water in the olive composition represents 40-50% of the pulp weight whereas the wash water for olive oil processing corresponds to 5% of processed olive weight and wash water equipment represent 5-10%. Therefore, wastes produced by traditional olive oil extraction system (discontinuous type) correspond to 50-60% of the processed drupe weight. In the case of continuous extraction systems the water used to thin out olive pastes during oil extraction has to be considered. The wastewater production has consequently increased and achieved values of 90-120% of the processed olive weight (Mounicif *et al.*, 1993).

These abundant volumes of wastewater have high values of Biological Oxygen Demand (BOD<sub>5</sub>) and Chemical Oxygen Demand (COD) because of their content of organic matter. So OOMWW produced during the olive oil extraction process, consists of an effluent liquid that possesses a considerable polluting charge and causes evident problems of disposal in the European areas of olive cultivation. OOMWW shows a low leaning towards biological treatments, which would allow for its disposal due to the consistent presence of organic substances including sugars, tannins, polyphenols, polyalcohols, pectins, and lipids (D'Annibale *et al.*, 1998). On the one hand, phenols are responsible of the high values of COD and BOD<sub>5</sub>, on the other hand if recovered, they might be reused as natural antioxidants as a replacement for BHA (2(3) tert-butyl-4-hydroxyanisole), BHT (3,5-di-tert-butyl-4-hydroxytoluene), TBHA (tert-butyl-hydroxyl-amine-acetate), NDGA (nord-hydroguaiaretic acid) and other synthetic antioxidants (ascorbic acid, gallate propyl, gallateottil, gallatedodecyle). Phenol recovery would result in a significant reduction in the COD and BOD<sub>5</sub> values of OOMWW. The phenolic composition of OOMWW has been studied in several recent works (Della Greca *et al.*, 2004; Obied *et al.*, 2005; Servili *et al.*, 1999). The phenolic composition of OOMWW is very different from that of the olive fruit. Olives are rich in secoiridoid glucosides whereas OOMWW showed a high concentration of secoiridoid derivatives such as hydroxytyrosol and the dialdehydic form of decarboxymethyl oleuropein aglycon. The OOMWW phenolic fraction is characterized by a great complexity, as demonstrated by Bianco *et al.* (2003); one of the most abundant and very interesting compounds from a nutritional point of view is hydroxytyrosol, which has been widely studied, demonstrating its antioxidant and beneficial health properties as well as its good bioavailability. Hydroxytyrosol scavenges free radicals (Visioli *et al.*, 1999) and inhibits human low-density lipoprotein (LDL) oxidation (Aruoma *et al.*, 1998; Casalino *et al.*, 2002; Deiana *et al.*, 1999). For economic reasons, OOMWW is usually concentrated in evaporation ponds and left to dry throughout the summer season (Borja *et al.*, 1992; Benitez *et al.*, 1997). The polluting power of olive oil mill wastewater depends on the high content of organic substances (sugars, pectins, lipids, nitrogen compound, alcohols and acids). Generally COD is between 100 and 190 g L<sup>-1</sup> of oxygen and the BOD<sub>5</sub> is 50-140 g L<sup>-1</sup> of oxygen. The polluting charge is principally due to the high concentrations of polyphenols (1-10 g L<sup>-1</sup>) that have different molecular weights, strong antimicrobial and phytotoxic properties and are also resistant to biological degradation.

The recovery of phenolic antioxidants by the waste seems achievable to produce substances industrially exploitable as supplemental food and the phytochemicals as chemical preservative compounds which can replace synthetic products. If the objective can be reached, significant economic gains can be

obtained olive oil production. Therefore, in the present work, a study of the recovery of phenolic fraction from the olive oil mill wastewater has been carried out because of its high antioxidant properties. The tests mainly revealed the viability of the extraction of phenols from wastewater and the recovery of the same fraction from the water previously concentrated. In addition, an evaluation of the antioxidant capacity has been successively conducted on several fractions obtained from differently treated wastewater.

## 2. MATERIALS AND METHODS

### 2.1. Materials

Fresh olive oil mill wastewater was supplied by the continuous three-phases of olive oil production process located in Reggio Calabria Province (Italy). The samples, collected in October 2009, were stored at room temperature before analyses. OOMWW was submitted to the following treatments: concentration at 50% of the initial volume in a rotary evaporator at 50 °C (OOMWW<sub>R</sub>) and concentration at 50% of the initial volume in an oven at 80 °C (OOMWW<sub>O</sub>). These tests were compared with the control (OOMWW).

### 2.2. Chemicals

Acetonitrile, methanol and acetic acid were HPLC-grade solvents purchased from Carlo Erba (Milan, Italy); Folin-Ciocalteu phenol reagent, sodium carbonate anhydrous, tyrosol, gallic acid, vanillic acid, *p*-coumaric acid, ferulic acid were obtained from Fluka (Buchs, Switzerland); oleuropeinglucoside was supplied from Extrasynthèse (Genay, France). 2,2 diphenyl-1-picrylhydrazyl (DPPH<sup>•</sup>) was obtained from Sigma-Aldrich Chemical Co. (St Louis, MO, USA).

### 2.3. Methods

The dry matter content was determined by oven drying at 105 °C up to a constant weight. The pH values were measured using a pH-meter (Crison basic 20, with a 50 cm electrode). These analyses were performed on three homogenized samples. The total polyphenols were extracted from olive oil mill wastewater following the Folin-Ciocalteu assay (Waterman and Mole, 1994): the samples were subjected to extraction with an ethylacetate/acetone solution (2:1, v/v) and then dehydration in a rotary evaporator and the residue was recovered in a 1 mL methanol/water solution (4:1, v/v). 2.5 mL of Folin-Ciocalteu's reagent and 5 mL of a 20% sodium carbonate solution were added to the phenolic extract in a 100 mL volumetric flask, filled to volume with deionized water and allowed to stand for 6 hours at room temperature and in the dark. The absorbance was measured at 725 nm using a Perkin-Elmer UV-Vis Lambda2 spectrometer. The total phenol concentrations were expressed as milligrams of

gallic acid equivalents per liter of fresh weight. Measurements were made in triplicate. To obtain extracts for HPLC analysis, 10 mL of olive oil mill wastewater, previously acidified to pH 2 with HCl, were diluted with 15 mL of hexane and then centrifuged at 3000 rpm for 5 min. The dilution was repeated twice, then the OOMWW was mixed with 10 mL of ethyl acetate and the mixture was vigorously shaken and centrifuged for 5 min at 3000 rpm. The phases were separated and the extraction was repeated four times. The organic extract dissolved in ethyl acetate was evaporated under vacuum at 40°C in a rotary evaporator. The residue was dissolved in 4 mL of methanol:water 1:1 and analyzed by HPLC. Qualitative and quantitative evaluations of phenolic compounds were conducted. The analyses were performed by an HPLC Knauer (Asi, Advanced Scientific Instruments, Berlin, Germany) smartline pump 1000, a UV Waters 486 detector coupled with a reversed phase C18 column, 120 Å, 4 m, 4.6 mm ID × 250 mm, 5 m particle size (Polymer Laboratories); guard column (4.6 mm ID, 5 m analytical particle size); 1 mL min<sup>-1</sup> flow; mobile phase: A = acetic acid : water solution (2:98, v:v) and B = methanol. The following gradient was used: 95% A and 5% B at the starting time, 70% A and 30% B at 25 min, 60% A and 40% B at 35 min, 52% A and 48% B at 40 min, 30% A and 70% B at 50 min, 100% B at 55 min. The eluents were detected at 280 nm at room temperature using gallic acid as internal standard (0.8 mg mL<sup>-1</sup>). The samples were filtered through a 0.45 μm membrane filter and 10 μL of this solution were injected into the column. The response factors were similar to those determined by Mateos *et al.* (2001). The antioxidant activity was measured according to the methodology described by Brand-Williams *et al.* (1995) based on the bleaching reaction of a stable free radical, 2,2-diphenyl-1-picrylhydrazyl (DPPH<sup>•</sup>) (Carlo Erba, MI, Italy) in the presence of the samples. The samples were left to react for 2 h 30 min in a cuvette containing 3 mL of a methanol solution 6·10<sup>-5</sup> M of DPPH<sup>•</sup>. Analysis was carried out at 515 nm wavelength and at ambient temperature, thus the risk of thermal degradation of the tested molecules is eliminated (Bondet *et al.*, 1997) to obtain a decrease in absorbance by the DPPH<sup>•</sup> radical (the decoloration curve of the radical follows very slow kinetics). A graph of absorbance vs. time shows that its decrease followed a 4<sup>th</sup> order kinetic ( $r^2 \geq 0.99$ ). Antioxidant capacity was expressed as  $-OD^{-3} \text{ min}^{-1} \text{ mL}^{-1}$  by the reaction rate  $k$  and calculated by the following equation:  $(1/A^3) - (1/A_0^3) = -3kt$  where  $A_0$  is the initial optical density and  $A$  is the optical density at increasing time  $t$ ,  $OD$  is optical density.

#### 2.4. Statistical analysis

A one-way variance analysis (ANOVA) was applied to the data to determine the significant differences ( $P \leq 0.05$ ). SPSS Software (Version 11.0, SPSS Inc., Chicago, IL, USA) was used for data processing.

### 3. RESULTS AND DISCUSSION

In Table 1, results of the pH values and the dry matter contents of the samples are reported. For the applied operations, the percentage of dry matter successively increased from 6.02 to 10.37 %, in the concentrated samples. The pH value did not significantly vary despite the treatments.

The total phenols were reported in Table 2 as mg of gallic acid equivalents (GAE) by a standard curve reference ( $y = 1.2699x - 0.0142$ ,  $r^2 = 0.9988$ ). The highest total phenol content (238.50 mg L<sup>-1</sup>) was evidently found in the fresh OOMWW extract, whereas, the original level of GAE decreased as consequence of the dehydration treatments and the successive oxidation reaction due to polyphenol oxidase (PPO) activity as reported in a previous work (Marsilio *et al.*, 2001). The total phenol levels determined in this way were not the absolute measurements of the amount of phenolic compounds but were in fact based on their reducing chemical capacity relative to an equivalent reducing capacity of gallic acid (Mc Donald *et al.*, 2001).

The identification of phenolic compounds in the OOMWW extracts was performed by HPLC-UV. The HPLC analysis confirmed hydroxytyrosol as the major representative phenolic compound as reported in literature (Fki *et al.*, 2005). In fact, its concentration ranged from 24.63 to 64.9 to mg L<sup>-1</sup> in the samples. Vanillic acid was the second represented one and tyrosol, ferulic acid, *p*-coumaric acid, 3,4 DHPEA-EA (the dialdehydic form of decarboxymethyl oleuro pein aglicon) and HPEA-EDA (the dialdehydic form of decarboxymethyl ligstroside aglicon) were present in lower amounts.

The different phenolic compounds responded similarly to the treatment of concentration. Their amount decreased after applying a high thermal process (80 °C) but, it increased significantly as a result of the rotary evaporator with a lower temperature. This could be considered to reduce the polyphenol content in the wastewater during a high thermal treatment, such as concentration in the oven. A general decrease in the single phenol quantity in dry matter was observed and revealed that the molecules showed a general degradation trend after the wastewater concentration. The percentage of the phenolic compounds followed different trends.

Table 1  
pH values and dry matter contents  
of different OOMWW

Samples	pH	Dry matter (%)
OOMWW	4.80 <sup>a</sup>	6.02 <sup>b</sup>
OOMWW <sub>R</sub>	4.78 <sup>a</sup>	10.37 <sup>a</sup>
OOMWW <sub>0</sub>	4.75 <sup>a</sup>	10.37 <sup>a</sup>
Sig.	n.s.	*

Data represent the mean of three replicates. Data followed by different letters are significantly different by Duncan's multiple range test. \*Significance at  $P \leq 0.05$ . \*\*Significance at  $P \leq 0.01$ , n.s. not significant.

Table 2  
Phenolic composition and total phenols of different OOMWW

	OOMWW			OOMWW <sub>R</sub>			OOMWW <sub>O</sub>			Sig.
	mgL <sup>-1</sup>	mgL d.m. <sup>-1</sup>	%	mgL <sup>-1</sup>	mgL d.m. <sup>-1</sup>	%	mgL <sup>-1</sup>	mgL d.m. <sup>-1</sup>	%	
Hydroxytyrosol	57.29 <sup>b</sup>	951.66	40.74	63.84 <sup>a</sup>	615.62	40.69	24.83 <sup>c</sup>	239.39	38.75	**
Tyrosol	14.52 <sup>b</sup>	241.11	10.32	21.02 <sup>a</sup>	202.65	13.39	11.27 <sup>c</sup>	108.68	17.59	**
Vanillic acid	52.64 <sup>a</sup>	874.34	37.43	52.68 <sup>a</sup>	507.96	33.57	18.20 <sup>b</sup>	175.46	28.4	**
Ferulic acid	8.75 <sup>b</sup>	145.27	6.22	9.81 <sup>a</sup>	94.6	6.25	4.78 <sup>c</sup>	46.05	7.45	**
<i>p</i> -Coumaric acid	5.41 <sup>b</sup>	89.78	3.84	6.47 <sup>a</sup>	62.34	4.12	4.90 <sup>c</sup>	47.25	7.65	*
3,4-DHPEA-EA	1.05 <sup>a</sup>	17.36	0.74	1.08 <sup>a</sup>	10.37	0.69	0.07 <sup>b</sup>	0.68	0.11	**
HPEA-EDA	0.90 <sup>b</sup>	14.95	0.64	1.50 <sup>a</sup>	14.46	0.96	0.13 <sup>c</sup>	1.21	0.2	**
Total phenols(GAE)	238.5 <sup>a</sup>	3961.79		162.26 <sup>b</sup>	1564.71		124.05 <sup>c</sup>	1196.24		**

Data represent the mean of three replicates. Data followed by different letters are significantly different by Duncan's multiple range test. \* Significance at  $P \leq 0.05$ . \*\* Significance at  $P \leq 0.01$ , n.s. not significant.

Although the ratio between the single phenols was generally constant, concentration in the oven seemed to increase the percentage values of tyrosol, ferulic acid and *p*-coumaric acid, whereas the concentration by rotavapor had the same effect only for the tyrosol and the HPEA-EDA content. In addition, hydroxytyrosol, vanillic acid and the 3,4-DHPEA-EA decreased proportionally after both thermal treatments.

As confirmed by Soleas (1997), antioxidant activity is highly correlated to the polyphenols, but not all phenolic compounds possess the same biological activity and their chemical compositions are strongly influenced by technological treatments as studied by Manzocco *et al.* (1998). The antioxidant compounds with molecular structures, identical to those of polyphenols, act to stop chain reactions (for example the oxidation) and form species of stable radicals after the elimination of a hydrogen atom. It is well known that many antioxidants can be significantly lost as a consequence of the processing as well as during prolonged storage (Johnson, 1991).

Nevertheless, it was recently demonstrated that thermal treatments can induce the formation of compounds with new and/or higher antioxidant properties (Nicoli *et al.*, 1997; Anese *et al.*, 1998). From the obtained results reported in Table 3, it is possible to denote that no concentrated OOMWW

had the lowest value of antioxidant capacity (44.3  $-\text{OD}^{-3} \cdot \text{min}^{-1} \cdot \text{mL}^{-1}$ ), whereas the OOMWW<sub>R</sub> demonstrated a higher radical scavenging property than OOMWW<sub>O</sub> (104.73 and 77.54  $-\text{OD}^{-3} \cdot \text{min}^{-1} \cdot \text{mL}^{-1}$ , respectively).

#### 4. CONCLUSIONS

The application of high concentration temperatures induced a strong reduction in total phenol content in OOMWW<sub>O</sub>, whereas in OOMWW<sub>R</sub> the low values of GAE could be ascribed to the oxidation by PPO activity which is prolonged under those thermal conditions (50 °C).

A polymerization of simple phenolic compounds probably took place because of the prolonged time of exposure to the processing temperature, as also observed in other studies (Pinelo, 2005). The concentration in the rotary evaporator most likely promoted the formation of polymers so that the OOMWW<sub>R</sub> sample showed the highest antioxidant property due to the cited achieved polymerization of polyphenols unlike the non thermal treated sample. Also, the OOMWW<sub>O</sub> sample has greater chain-breaking activity than the OOMWW, but this result could not be ascribable to the phenolic contents. During thermal treatments, besides the nutrient loss, foods can be subjected to other chemical changes such as those resulting from the Maillard reaction. This reaction, which occurs when sugars condense with free amino acids, peptides or proteins, leads to the formation of a wide variety of intermediate products (Maillard reaction products, MRPs) that can act as antioxidants (Nicoli *et al.*, 1997). The waste water, as before mentioned, is rich in organic substances, such as sugars, proteins and polyphenols. These can take part in the Maillard reactions as they are formed during oxidative reactions. So, the loss of natural antioxidants in the heating process could be minimized or compensated by the formation of non-nutrient antioxidants such as MRPs, as demonstrated in other recent

Table 3  
Chain breaking activity of OOMWW

Samples	Chain breaking activity ( $-\text{OD}^{-3} \cdot \text{min}^{-1} \cdot \text{mL}^{-1}$ )
OOMWW	44.3 <sup>c</sup>
OOMWW <sub>R</sub>	104.73 <sup>a</sup>
OOMWW <sub>O</sub>	77.54 <sup>b</sup>
Sig.	**

Data represent the mean of three replicates. Data followed by different letters are significantly different by Duncan's multiple range test. \* Significance at  $P \leq 0.05$ . \*\* Significance at  $P \leq 0.01$ , n.s. not significant.

experiments (Manzocco *et al.*, 2001; Madrau *et al.*, 2009). In our study, the treatment by concentration of olive oil wastewater in a rotary evaporator and in the oven, involved an increase in its efficiency, so that this type of waste seems to be fit for a recovery and a successive reuse.

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