

1 *"This is the peer reviewed version of the following article: Biondi, A., Campolo, O., Desneux, N., Siscaro, G.,*
2 *Palmeri, V. and Zappalà, L., 2015. Life stage-dependent susceptibility of *Aphytis melinus* DeBach*
3 *(Hymenoptera: Aphelinidae) to two pesticides commonly used in citrus orchards. *Chemosphere*, 128,*
4 *pp.142-147..], which has been published in final doi [<https://doi.org/10.1016/j.chemosphere.2015.01.034>].*
5 *The terms and conditions for the reuse of this version of the manuscript are specified in the publishing*
6 *policy. For all terms of use and more information see the publisher's website"*

7

8 Life stage-dependent susceptibility of *Aphytis melinus* DeBach (Hymenoptera: Aphelinidae) to
9 two pesticides commonly used in citrus orchards

10

11 Antonio Biondi a , Orlando Campolo b , Nicolas Desneux c , Gaetano Siscaro a , Vincenzo Palmeri b , Lucia
12 Zappalà a

13 a University of Catania, Dipartimento di Agricoltura, Alimentazione e Ambiente, via Santa Sofia 100, 95123
14 Catania, Italy

15 b University “Mediterranea” of Reggio Calabria, Dipartimento di AGRARIA, Feo di Vito, 89122 Reggio
16 Calabria, Italy

17 c French National Institute for Agricultural Research (INRA), Institut Sophia Agrobiotech, 400 Route des
18 Chappes, 06903 Sophia-Antipolis, France

19

20 **highlights**

- 21· Lethal and sublethal effects of two pesticides were studied on *Aphytis melinus* .
- 22· Significant differences in parasitoid instars susceptibility were recorded.
- 23· Pesticide risk assessment and choice should include life-stage susceptibility.
- 24· The natural origin of the mineral oil is not guarantee of non-target safety.
- 25· Careful integration of *A. melinus* and the tested insecticides is recommended.

26 **ABSTRACT**

27 The assessment of pesticides compatibility with natural enemies is recommended before including
28 agrochemicals in integrated and organic pest management schemes. The lethal and sublethal effects of a
29 mineral oil and a juvenile hormone mimic (pyriproxyfen), on adults and larvae of *Aphytis melinus* , a key
30 ectoparasitoid of armored scale insect pests of citrus, such as *Aonidiella aurantii* , were evaluated. Mineral
31 oil caused very high mortality on the adults, while a lower acute toxicity was recorded on young instars. No
32 significant effects on their reproduction capacity and on the sex-ratio of the progeny were observed.
33 Pyriproxyfen had neither lethal nor sublethal effects (in terms of survived female fertility) on *A. melinus*
34 adults. However, parasitoid larvae exposed to this insecticide suffered strong acute toxicity and fertility
35 reduction (progeny number and proportion of female progeny). When adults were offered the choice to
36 parasitize treated and untreated scales they significantly preferred the control ones, and when they were
37 exposed to only treated scaled the parasitism rate was significantly lower only with mineral oil-treated hosts.
38 The significant differences in the susceptibility of the two parasitoid instars highlight the importance of
39 including this aspect in pesticide risk assessment procedures and in the choice of the pesticide and of the
40 treatment timing in the field. Overall, the results indicate potential for integrating *A. melinus* , both naturally
41 present and artificially released, and these insecticides only by appropriate timing of insecticide spraying and
42 parasitoid releases.

43 **INTRODUCTION**

44 The California red scale, *Aonidiella aurantii* (Maskell) (Hemiptera: Diaspididae), is an invasive pest of citrus
45 and despite its common name it is believed to have Asiatic origin (Compere, 1961). Nowadays, this species is
46 considered one of the most important pests of citrus in the Mediterranean basin and in several other citrus
47 growing areas worldwide (Jacas et al., 2010; Zappalà, 2010). The ectoparasitoid *Aphytis melinus* DeBach
48 (Hymenoptera: Aphelinidae) is a key natural enemy of *A. aurantii* (Compere, 1961; Siscaro et al., 1999;
49 Sorribas and Garcia-Marí, 2011) and it is mass reared for inoculative and augmentative releases in citrus

50 orchards (Moreno and Luck, 1992; Zappalà, 2010). The effectiveness of *Aphytis* spp. in the scale control could
51 depend on many agro-ecological factors, such as the fitness of the released insects (Vasquez and Morse,
52 2012), their field dispersal capacity (Zappalà et al., 2012), the availability of the susceptible host instars and
53 their size (Luck and Podoler, 1985), the interaction with other natural enemies (Heimpel et al., 1997a; Borer
54 et al., 2003; Vanaclocha et al., 2013a), and the presence in the tree canopy of insecticide residues (Suma et
55 al., 2009; Garcerá et al., 2013; Vanaclocha et al., 2013b). Although various environmentally friendly pest
56 control tools, such as habitat management, cultural practices, mating disruption, and biological control, are
57 currently adopted and implemented in citrus crops worldwide (Lim et al., 2006; Vacas et al., 2012; Aguilar-
58 Fenollosa and Jacas, 2013; Chueca et al., 2013), chemical treatments are frequently applied in citrus groves
59 (Rill et al., 2008; Garcerá et al., 2014; Monzò et al., 2014). The use of selective pesticides is therefore crucial
60 for maintaining natural enemy populations and their ecosystem services (Prabhaker et al., 2007; Suma et al.,
61 2009). Thus, the correct evaluation of potential side effects of pesticides on non-target organisms, such as
62 the natural enemies employed in citrus IPM and organic packages, is particularly crucial (Garcerá et al., 2013;
63 Planes et al., 2013). Pesticides could have both lethal (i.e. acute toxicity) and sublethal (i.e. influence on
64 various behavioral and physiological traits) effects on the exposed natural enemies (Desneux et al., 2007;
65 Biondi et al., 2012a). A comprehensive evaluation of undesired effects of agrochemicals should be therefore
66 based not only on short-term acute toxicity tests, but also on the assessment of physiological and behavioral
67 sublethal effects. These should include the potential long-term effects on natural enemy population
68 dynamics and their ecosystem services (He et al., 2012; Biondi et al., 2013; Saber and Abedi, 2013; Bengochea
69 et al., 2014). Moreover, most of the tests used to determine the side effects of pesticides on natural enemies
70 are performed on only a single developmental stage, primarily the adults, which are considered to be the
71 most exposed life stage. Immature developmental stages, however, may also be affected by pesticides, even
72 though they are usually concealed within the host (Schneider et al., 2004). In this framework, studying the
73 potential effects of pesticides on different developmental stages of the natural enemies could be relevant,
74 also considering their variable susceptibility which has been often highlighted, particularly when testing
75 Insect Growth Regulator (IGR) compounds (Ishaaya and Horowitz, 1995; Hoddle et al., 2001; Schneider et al.,
76 2004). Mindful of this context, we aimed at evaluating the compatibility of a narrow range mineral oil and a
77 juvenile hormone mimic with *A. melinus* when applied on adults and young instars of the parasitoid.
78 Laboratory bioassays were conducted to assess the acute toxicity and the sublethal effects on reproduction
79 on adults and young instars, as well as *A. melinus* parasitization activity on treated hosts.

80 **Material and methods**

81 **Insects**

82 A parthenogenetic strain of the oleander scale, *Aspidiotus nerii* Bouché (Hemiptera: Diaspididae), was reared
83 on organically grown squashes, *Cucurbita maxima* Duch. var. Butternut. Infested squashes were used for the
84 parasitoid rearing, following *A. lingnanensis* rearing methodology as described by DeBach and White (1960),
85 and modified by Raciti et al. (2003). All the insects used in this study were reared and kindly provided by the
86 Sicilian Regional Insectary (Biofabbrica Insetti Utili, Ramacca, Catania, Italy). In particular *A. nerii*-infested and
87 *A. nerii*-infested and parasitized squashes were promptly transferred to the laboratory, when needed for the
88 toxicological bioassays.

89 **Pesticides**

90 We evaluated under laboratory conditions the effects of two agrochemicals. The first was the narrow range
91 mineral oil Biolid E " (Emulsifiable Concentrate 80% a.s., Sipcam, Italy), generally used as adjuvant and/or
92 alone to control mites and scale pests when at the first instars on many different crops under conventional,
93 integrated and organic pest management programs (Biondi et al., 2012a). The other was pyriproxyfen, a
94 juvenile hormone mimic [Admiral 10EC" (Emulsifiable Concentrate 10.86% a.s.), Sumitomo Chemical, Japan],
95 which inhibits metamorphosis and embryogenesis in several insects (Ishaaya and Horowitz, 1995). Both

96 chemicals were applied at their maximum label dose: 2 L hL⁻¹ and 75 mL hL⁻¹ respectively. Untreated
97 controls were sprayed with tap water only. All the bioassays were conducted at 23 ± 1 °C, 60 ± 10% R.H. and
98 a photoperiod of 16:8 (L:D).

99 Acute toxicity on *A. melinus* adults

100 The acute toxicity was assessed exposing newly emerged (24–48 h) adults to freshly dried pesticide residues
101 on glass plates. Uniform deposits (1.3–1.8 mg/cm²) of pesticide solution or of water were obtained using a
102 Potter Precision Spray Tower (Burkard Manufacturing Co. Ltd.) to spray 6 glass plates (9 # 9 cm).
103 Subsequently, the plates were left into a chemical hood for about 2 h to complete drying and then assembled
104 to put together a cube (Suma et al., 2009). To avoid fumigation effect, each set of experimental arenas was
105 provided with a electric pump (Air fizz 100, Ferplast Spa, Italy) ensuring a constant 36 mL # min⁻¹ air-flow
106 (Suma et al., 2009). To mimic the field scenario where parasitoid adults use external nutrients (Beltrá et al.,
107 2013), a solution of pure honey and water (1:1) was provided in the experimental arena through a cotton
108 ball imbibed of honey solution. Ten parasitoids (five females and five males) were introduced in the arena
109 and left in contact with the pesticide residues for 24 h and then their survival was recorded. Adults were
110 considered dead if they did not react after being touched by a fine paint brush. Ten replicates per each
111 treatment were carried out.

112

113 Acute toxicity on *A. melinus* larvae

114 Bioassays on *A. melinus* juveniles were conducted using squashes infested with 1000 ± 100 oleander scale
115 third instar nymphs, i.e. the parasitoid preferred host instar (Rosen and DeBach, 1979). Infested squashes
116 were exposed to 10 females and 10 males of the parasitoid for 48 h in ventilated plastic boxes (35 # 25 # 45
117 cm) (l # w # h), provided with honey droplets. Two similar squashes infested with 500 third instars *A. nerii*
118 each, were simultaneously exposed to *A. melinus* adults per each replicate and treatment. In order to expose
119 *A. melinus* third instars to the tested insecticides or to the control solution, only one squash was sprayed
120 until run-off [using a 2 L aerosol hand sprayer (Matabi[®], Antzuola, Guipuzcoa, Spain)] 5 d after oviposition
121 initiation (Raciti et al., 2003). While, the other infested squash was moved to a different box kept under the
122 same environmental conditions. This second set of squashes provided a specific control data on parasitoid
123 emergence per each sprayed squash. The number and the sex-ratio of the emerged progeny from all the
124 squashes were scored daily for one week after the juvenile development completed, i.e. 12 d at these
125 environmental conditions (Raciti et al., 2003). The experiment was replicated ten times. The percentage of
126 young instars mortality was estimated as: $\frac{N_{cx} - N_{tx}}{N_{cx}} \times 100$ where N_{cx} is the number of *A. melinus*
127 adults emerged in the control squash c_x and N_{tx} is the number of emerged adults from the corresponding
128 treated squash t_x .

129 Sublethal effects on reproduction

130 The adults that survived the 24-h exposure to pesticide dry residue within the glass cubes, were placed for
131 one week with new hosts susceptible to be parasitized (third instar nymphs), at the rate of 100 scales per
132 parasitoid female. Survived parasitoids were released in couples (one male and one female (Pan et al., 2014))
133 inside plastic cylindrical isolators (45 cm³) fixed on infested squashes using a strip of modeling paste (Jovi[®],
134 Barcelona, Spain). The isolators were ventilated and supplied with honey droplets. The reproductive capacity
135 of the adults developed from topically exposed larvae (those coming from the “Acute toxicity on *A. melinus*
136 larvae” experiment) was assessed by placing a newly emerged couple for one week with new susceptible
137 hosts (at the density of 100/female), inside isolators similar to those described above for the adults survived
138 to the residual exposure to the pesticides. The number and the sex-ratio of the emerged progeny were scored
139 daily for one week starting 12 d after the oviposition beginning. All the trials were replicated 10 times.

140 Parasitism on treated hosts: choice and no-choice tests

141 In order to assess whether or not *A. melinus* is able to parasitize treated hosts on a treated substrate, the
142 parasitization activity of adult wasps on hosts previously treated with mineral oil, pyriproxyfen or water was
143 assessed. The squashes were left in a chemical hood for about 2 h to complete drying. Then, newly emerged
144 (624 h) *A. melinus* adults were released, in groups of 20 couples, on 2000 *A. nerii* third instars (100
145 hosts/female), and left for 48 h in a dual-choice and in a no-choice test. For both kinds of test, two squashes
146 infested with 1000 scales were exposed to the parasitoids for 48 h into the plastic boxes previously described
147 for the acute toxicity test on young instars. In the choice test one treated and one untreated infested squash
148 were placed in the same plastic box and exposed to *A. melinus* adults, while in the no-choice test, two treated
149 and two untreated infested squashes were placed in two different boxes and exposed to 20 *A. melinus* adults
150 per each box. Forty-eight hour after the end of the parasitization, all the scales were observed under the
151 binocular to assess the parasitism rate, i.e. the portion of parasitized scales (with at least one *A. melinus* egg
152 or larva) on the total exposed scales, was calculated. Each test and combination was replicated five times

153 Data analyses

154 Each dataset (adults and young instars) and the parasitism rates obtained in the no-choice test, were first
155 tested for homoscedasticity (Levene test) and normality (Shapiro–Wilk test) of their distribution and
156 transformed when necessary. Then, they were subjected to one-way ANOVA and Fisher’s Least Significant
157 Difference (LSD) test was used to determine which insecticidal treatments were significantly different at the
158 $P < 0.05$ level. The parasitism rates obtained in the choice test were analyzed as follows: the null hypothesis
159 that *A. melinus* parasitization choice was not affected by treatments (a response equal to 50:50 of parasitized
160 scales) was subjected to a chi-square goodness of fit test. All statistical analyses were conducted with
161 STATISTICA 7.0 (StatSoft Inc., OK, USA). Lethal (mortality) and sublethal effects (fertility output and
162 proportion of female progeny) of the tested pesticides were summarized in a reduction coefficient (Ex),
163 calculated for adults and young instars separately, according to the formula described by Biondi et al. (2012a).
164 These values were then compared to the standards for laboratory ecotoxicological tests of the International
165 Organization for Biological Control (IOBC) which include four categories: (1) harmless: $E < 30\%$, (2) slightly
166 harmful: $30\% < E < 80\%$, (3) moderately harmful: $80\% < E < 99\%$, and (4) harmful: $E > 99\%$ (Sterk et al., 1999).

167 Results

168 Acute toxicity. The acute toxicity trials showed a significant effect of the treatment both on the adults ($F =$
169 69.14 ; $df = 2, 23$; $P < 0.001$) and on the young instars ($F = 11.24$; $df = 2, 48$; $P < 0.001$). Mineral oil killed most
170 of the tested adult wasps ($97.2 \pm 1.9\%$), and all the females, making impossible the effects on the progeny
171 evaluation. Pyriproxyfen resulted less toxic towards adults ($38.4 \pm 5.6\%$), although the mortality was
172 significantly higher than that registered for the control (Table 1). The highest percentage of estimated young
173 instars mortality was obtained from the parasitized scales sprayed with pyriproxyfen ($37.0 \pm 6.1\%$), which
174 resulted significantly higher than the mortality of both scales treated with mineral oil ($13.2 \pm 3.6\%$) and that
175 of the control (Table 1). No significant differences ($F = 0.45$; $df = 2, 48$; $P = 0.637$) were found in the sex-ratio
176 of the adults emerged from the treated parasitized scales (Table 2).

177 Sublethal effects on reproduction

178 Pyriproxyfen did not significantly affect either the fertility of the adult females that survived the exposure to
179 its residues (42.8 ± 11.1 offspring/female; $F = 3.67$; $df = 1, 18$; $P = 0.092$) (Table 1) or the sex ratio of their
180 progeny ($F = 0.35$; $df = 1, 18$; $P = 0.57$) (Table 2). While, the trials conducted on *A. melinus* young instars
181 showed a significant effect of the treatment in terms of progeny production of the developed individuals ($F =$
182 3.75 ; $df = 2, 28$; $P < 0.001$). Indeed, the mean fertility of the adults developed from the young instars treated
183 with pyriproxyfen was 14.2 ± 4.1 progeny/female and it significantly differed from mineral oil (30.9 ± 5.4) and
184 the control (31.1 ± 5.9) (Table 1). Besides, pyriproxyfen significantly disrupted the sex ratio of the progeny

185 produced by the specimens treated as larvae ($F = 6.55$; $df = 2, 28$; $P < 0.001$), causing a reduction in the female
186 proportion (Table 2).

187 Parasitism on treated hosts

188 In the dual-choice tests a significantly lower parasitism rate on the hosts treated with both pesticides (mineral
189 oil: $19.2 \pm 4.5\%$; pyriproxyfen: $29.6 \pm 5.3\%$) compared to the control ($61.6 \pm 2.6\%$ and $43.2 \pm 1.62\%$,
190 respectively) was recorded (Fig. 1). In the nochoice test the effect of the treatment was also significant ($F =$
191 7.09 ; $df = 2, 14$; $P < 0.01$). However, the parasitism rate was significantly lower (LSD test $P < 0.05$) only on
192 hosts treated with mineral oil ($26.5 \pm 9.9\%$), compared with pyriproxyfen ($63.2 \pm 9.9\%$) and with the control
193 ($82.7 \pm 9.1\%$). Treated scales were observed and no significant difference in mortality between treated and
194 control scales was found.

195 Reduction coefficient (Ex) and IOBC toxicity classes

196 The calculated reduction coefficient was 96.81% for the mineral oil and 42.22% for pyriproxyfen in the trials
197 on lethal and sublethal effects on adults. According, pesticides were classified as moderately harmful (class
198 3) and slightly harmful (class 2) to adults, respectively. By contrast, the reduction coefficient estimated for the
199 impact of the two pesticides on the wasp young instars was 16.55% for mineral oil, which was classified as
200 harmless, and 81.03% for pyriproxyfen, then classified as moderately harmful.

201 Discussion

202 We used the squash/*A. nerii*/*A. melinus* laboratory system, to study how two agrochemicals, commonly used
203 in citrus agro-ecosystems, can affect the population dynamics and the biocontrol activity of a parasitoid of a
204 citrus key pest. The obtained results showed differences in the susceptibility of the two parasitoid
205 developmental stages to the two pesticides. In particular, the narrow range mineral oil was lethal to *A.*
206 *melinus* adults through residual contact on inert substrate, and it reduced the parasitization activity of the
207 females when treated hosts were offered. Meanwhile, mineral oil did not show any toxicity on the parasitoid
208 young instars when applied on parasitized scales. Interestingly, we found a divergent toxicological behavior
209 of pyriproxyfen towards the two parasitoid life stages. Pyriproxyfen slightly affected *A. melinus* adult survival
210 and did not reduce the reproduction of the adults survived to that exposure, but it strongly impacted the
211 juvenile survival and especially the reproduction capacity (both progeny size and sex-ratio) of the adults that
212 developed from the treated parasitized hosts. Moreover, when treated and untreated hosts were exposed
213 simultaneously, parasitoids always preferred parasitizing the control hosts. While, when *A. melinus* was not
214 given any choice, this trend was consistent only for the mineral oil-treated hosts on which the parasitization
215 rate was lower than on untreated hosts. Although the parasitization reductions caused by both insecticides
216 were not very high numerically, this decrease could be ecologically important for its effects on the parasitoid
217 biocontrol services, including the recolonization capacity in a treated environment. The obtained results
218 could be explained by the different mode of action of the two pesticides. In particular, the mineral oil was
219 toxic to *A. melinus* adults as contacticide, most likely blocking the breathing pores and/or the insect cuticle,
220 as in other minute and soft-bodied arthropods after residual or topical contact (NajarRodriguez et al., 2007).
221 Similarly, a detrimental effect was noted on the parasitization activity of *A. melinus* on sprayed hosts both
222 in the choice and no-choice experiment, which could decrease the recolonization ability of the parasitoid in
223 treated environments (Desneux et al., 2006; Biondi et al., 2013). These data confirm those obtained in
224 previous trials where the same parasitoid was highly susceptible to mineral oil fresh residue on glass surfaces
225 (Suma et al., 2009) and on freshly sprayed citrus leaves for 24 h (Campbell, 1975). While, Vanaclocha et al.
226 (2013b) found a lower (27%) *A. melinus* mortality after exposing parasitoid adults to petroleum oil in the
227 laboratory. These divergent results may be due to the fact that Vanaclocha et al. (2013b) used another
228 commercial product, tested at lower concentration (1.5 vs 2.0 L hL⁻¹), and gaveto the parasitoids the
229 opportunity to avoid the insecticide exposure and/or to recover in untreated areas of the experimental
230 arena. This latter point, although it should be further investigated through behavioral observations, could

231 have influenced the parasitoid survival significantly. Meanwhile, in the present study we provide for the first
232 time evidences on the mineral oil safety towards both the parasitoid larvae feeding under the scale waxy
233 cover, and the developed adults that did not suffer any potential exposure to the oil residues. In contrast,
234 the injuriousness towards immature stages of pyriproxyfen was shown by the low emergence rates that we
235 recorded from parasitized hosts sprayed with this IGR. To explain this important pyriproxyfen impact on
236 young instars, we suggest three hypotheses: (i) the insecticide was able to penetrate into the scale cover and
237 contaminated, directly by contact and/or indirectly by ingestion of contaminated hosts, the parasitoid young
238 instars causing molt and/or metamorphosis disorders that affected their survival (Ishaaya and Horowitz,
239 1995; Planes et al., 2013); (ii) pyriproxyfen did not penetrate the scale cover, but newly formed adults
240 ingested insecticide residues when chewing their way through the scale cover during emergence and caused
241 behavioral impairment that prevented the successful parasitoid emergence, as already recorded for moth
242 larvae emerging from pyriproxyfen-treated eggs (Tomé et al., 2012); and (iii) a combination of these two
243 phenomena was responsible of the low emergence rates. Nevertheless, the results obtained by Rill et al.
244 (2008), clearly showed that pyriproxyfen and another IGR insecticide, buprofezin, were not toxic towards *A.*
245 *melinus* eggs and larvae, when California red scale infested lemons were dipped into the insecticide
246 solutions. These divergent results could be explained by (i) the differential permeability of the cover of the
247 two scale species employed in the two experiments, both intrinsic of the scales or induced by the two rearing
248 vegetal hosts, i.e. lemons and squashes, as showed in the case of spinosad susceptibility of egg-parasitoid
249 pupae into the eggs of various insect species (Biondi et al., 2012b) and (ii) or to the intraspecific variation in
250 insecticide resistance that has been previously reported in *A. melinus* (Rosenheim and Hoy, 1986, 1988). The
251 intraspecific variation in pyriproxyfen susceptibility of this parasitoid species may also explain the discrepancy
252 between our results and those of Gonzalez-Zamora et al. (2013) that obtained 100% of mortality when
253 exposing for 24 h adults to pyriproxyfen residues on inert substrate. However, in Vanaclocha et al. (2013b)
254 the mortality of *A. melinus* adults was not significant when parasitoids were exposed to dry residues on
255 leaves in the laboratory. Besides, this compound caused a reduction in the fertility of the adults that survived
256 the juvenile exposure and emerged from the treated young instars. This could be due to the fact that juvenile
257 hormone analogs, such as pyriproxyfen, are transported to the ovary or they penetrate into the eggs where
258 they block embryogenesis at blastokinesis (Staal, 1975; Retnakaran et al., 1985). This would explain why the
259 reduction in fertility did not occur when the biological instars exposed to the IGR residue is the adult one.
260 Indeed, although *A. melinus* is a synovigenic parasitoid with an ovigeny index of 0 to 0.07 (Heimpel et al.,
261 1997b), it may have already developed eggs before the pesticide exposure, i.e. 1–2 d after emergence, while
262 pyriproxyfen may have blocked the ovigeny process of young adults that developed from treated young
263 instars. Sex ratio was affected (higher male proportion) only in the progeny of adults exposed to pyriproxyfen
264 as immatures. Considering that arrhenotokous parthenogenesis (i.e. unfertilized females produce only males)
265 occurs in *A. melinus* (Rosen and DeBach, 1979), we can argue that higher male progeny may be due to male
266 sterility or mating behavior alterations.

267 **Conclusion**

268 Overall, this research brings useful findings for optimizing integrated and organic management programs
269 against insect scale pests of citrus. The results suggest a careful timing of the applications of these two
270 insecticides when thought to be integrated with *A. melinus* field activity. In order to combine mineral oil
271 applications and *A. melinus* augmentative releases, the latter should be performed before the treatments or
272 when the mineral oil residues are degraded below a toxicity threshold for *A. melinus* in citrus orchards, i.e.
273 one week after the treatment, as suggested by Campbell (1975). In contrast, pyriproxyfen applications should
274 be strictly avoided when parasitoids are developing into their hosts. Then, all these findings stress the
275 importance of pest and natural enemy monitoring for the optimization of pest control programs in citrus
276 groves (Zappalà, 2010; Campolo et al., 2014; Monzò et al., 2014). The detrimental results obtained exposing
277 the parasitoid to the narrow range mineral oil residues stress the idea that naturally derived pesticides, thus
278 authorized in organic farming, are not necessarily safe for non-target organisms (Biondi et al., 2012a, 2012b,

279 2013; de Castro et al., 2013; Martinou et al., 2014), and that the origin of a given compound could not really
280 relate to its toxicological properties in cropping conditions. Moreover, from the risk assessment perspective
281 all these results stress how important is the accurate toxicity evaluation on both lethal and sublethal effects
282 toward multiple developmental stages. Besides, the data on the fertility of emerged adults would suggest
283 continuing the evaluation of potential transgenerational effects of pyriproxyfen, being these crucial on the
284 demography of survived individuals (Ali et al., 2012; Liang et al., 2012).

285 **ACKNOWLEDGMENTS**

286 The authors thank the Sicilian Regional Insectary (Biofabbrica Insetti Utili, Ramacca, Catania, Italy) for
287 providing the insects used in the experiments.

288 **REFERENCES**

- 289 Aguilar-Fenollosa, E., Jacas, J.A., 2013. Effect of ground cover management on Thysanoptera (thrips) in
290 clementine mandarin orchards. *J. Pest Sci.* 86, 469–481.
- 291 Ali, A., Ahmad, F., Biondi, A., Wang, Y., Desneux, N., 2012. Potential for using *Datura alba* leaf extracts against
292 two major stored grain pests, the khapra beetle *Trogoderma granarium* and the rice weevil *Sitophilus oryzae*.
293 *J. Pest Sci.* 85, 359–366.
- 294 Beltrá, A., Tena, A., Soto, A., 2013. Reproductive strategies and food sources used by *Acerophagus n.* sp. near
295 *coccos*, a new successful parasitoid of the invasive mealybug *Phenacoccus peruvianus*. *J. Pest Sci.* 86, 253–
296 259.
- 297 Bengochea, P., Budia, F., Viñuela, E., Medina, P., 2014. Are kaolin and copper treatments safe to the olive
298 fruit fly parasitoid *Psytalia concolor*? *J. Pest Sci.* 87, 351–359.
- 299 Biondi, A., Desneux, N., Siscaro, G., Zappalà, L., 2012a. Using organic-certified rather than synthetic pesticides
300 may not be safer for biological control agents: selectivity and side effects of 14 pesticides on the predator
301 *Orius laevigatus*. *Chemosphere* 87, 803–812.
- 302 Biondi, A., Mommaerts, V., Smagghe, G., Viñuela, E., Zappalà, L., Desneux, N., 2012b. The non-target impact
303 of spinosyns on beneficial arthropods. *Pest Manage. Sci.* 68, 1523–1536.
- 304 Biondi, A., Zappalà, L., Stark, J.D., Desneux, N., 2013. Do biopesticides affect the demographic traits of a
305 parasitoid wasp and its biocontrol services through sublethal effects? *Plos One* 8 (9), e76548.
- 306 Borer, E.T., Briggs, C.J., Murdoch, W.W., Swarbrick, S.L., 2003. Testing intraguild predation theory in a field
307 system: does numerical dominance shift along a gradient of productivity? *Ecol. Lett.* 6, 929–935.
- 308 Campbell, M.M., 1975. Duration of toxicity of residues of malathion and spray oil on citrus foliage in South
309 Australia to adults of a California Red Scale parasite *Aphytis melinus* DeBach (Hymenoptera: Aphelinidae).
310 *Aust. J. Entomol.* 14, 161–164.
- 311 Campolo, O., Malacrinò, A., Laudani, F., Maione, V., Zappalà, L., Palmeri, V., 2014. Population dynamics and
312 temperature-dependent development of *Chrysomphalus aonidum* (L.) to aid sustainable pest management
313 decisions. *Neotrop. Entomol.* 43, 453–464.
- 314 Chueca, P., Garcerá, C., Urbaneja, A., Moltó, E., 2013. A new mechanized cultural practice to reduce *Ceratitidis*
315 *capitata* Wied. populations in area-wide IPM. *Span. J. Agric. Res.* 11, 1129–1136.
- 316 Compere, H., 1961. The red scale and its insect enemies. *Hilgardia* 31, 173–278. de Castro, A.A., Corrêa, A.S.,
317 Legaspi, J.C., Guedes, R.N.C., Serrão, J.E., Zanoncio, J.C., 2013. Survival and behavior of the insecticide-
318 exposed predators *Podisus nigrispinus* and *Supputius cincticeps* (Heteroptera: Pentatomidae). *Chemosphere*
319 93, 1043–1050.

- 320 DeBach, P., White, E.B., 1960. Commercial mass culture of the California red scale parasite. Calif. Agric. Expt.
321 Sta. Bull., 770
- 322 Desneux, N., Ramirez-Romero, R., Kaiser, L., 2006. Multistep bioassay to predict recolonization potential of
323 emerging parasitoids after a pesticide treatment. Environ. Toxicol. Chem. 25, 2675–2682.
- 324 Desneux, N., Decourtye, A., Delpuech, J.M., 2007. The sublethal effects of pesticides on beneficial arthropods.
325 Annu. Rev. Entomol. 52, 81–106.
- 326 Garcerá, C., Ouyang, Y., Scott, S.J., Moltó, E., Grafton-Cardwell, E., 2013. Effects of spirotetramat on *Aonidiella*
327 *aurantii* (Homoptera: Diaspididae) and its parasitoid, *Aphytis melinus* (Hymenoptera: Aphelinidae). J. Econ.
328 Entomol. 106, 2126–2134.
- 329 Garcerá, C., Moltó, E., Chueca, P., 2014. Factors influencing the efficacy of two organophosphate insecticides
330 in controlling California red scale, *Aonidiella aurantii* (Maskell). A basis for reducing spray application volume
331 in mediterranean conditions. Pest Manage. Sci. 70, 28–38.
- 332 Gonzalez-Zamora, J.E., Castillo, M.L., Avilla, C., 2013. Side effects of different pesticides used in citrus on the
333 adult stage of the parasitoid *Aphytis melinus* DeBach (Hymenoptera Aphelinidae) and its progeny. Span. J.
334 Agric. Res. 11, 494–504.
- 335 He, Y.X., Zhao, J.W., Zheng, Y., Desneux, N., Wu, K., 2012. Lethal effect of imidacloprid on the coccinellid
336 predator *Serangium japonicum* and sublethal effects on predator voracity and on functional response to the
337 whitefly *Bemisia tabaci*. Ecotoxicology 21, 1291–1300.
- 338 Heimpel, G.E., Rosenheim, J.A., Mangel, M., 1997a. Predation on adult *Aphytis* parasitoids in the field.
339 Oecologia 110, 346–352.
- 340 Heimpel, G.E., Rosenheim, J.A., Kattari, D., 1997b. Adult feeding and lifetime reproductive success in the
341 parasitoid *Aphytis melinus*. Entomol. Exp. Appl. 83, 305–315.
- 342 Hoddle, M.S., van Driesche, R.G., Lyon, S.M., Sanderson, J.P., 2001. Compatibility of insect growth regulators
343 with *Eretmocerus eremicus* (Hymenoptera: Aphelinidae) for whitefly (Homoptera: Aleyrodidae) control on
344 poinsettias. 1. Laboratory assays. Biol. Control 20, 122–131.
- 345 Ishaaya, I., Horowitz, A.R., 1995. Pyriproxyfen, a novel insect growth regulator for controlling whiteflies:
346 mechanisms and resistance management. Pestic. Sci. 43, 227–232.
- 347 Jacas, J., Karamaouna, F., Vercher, R., Zappalà, L., 2010. Citrus pest management in the northern
348 mediterranean basin (Spain, Italy and Greece). In: Ciancio, A., Mukerji, K.G. (Eds.), Integrated Management of
349 Arthropod Pests and Insect Borne Diseases. Springer, pp. 3–26.
- 350 Liang, P., Tian, Y.A., Biondi, A., Desneux, N., Gao, X.W., 2012. Short-term and transgenerational effects of the
351 neonicotinoid nitenpyram on susceptibility to insecticides in two whitefly species. Ecotoxicology 21, 1889–
352 1898.
- 353 Lim, U., Zappalà, L., Hoy, M.A., 2006. Pre-release evaluation of *Semielacher petiolatus* (Hymenoptera:
354 Eulophidae) in quarantine for the control of citrus leafminer: host discrimination, relative humidity, and
355 alternative hosts. Biol. Control 36, 65–73.
- 356 Luck, R.F., Podoler, H., 1985. Competitive exclusion of *Aphytis lingnanensis* by *Aphytis melinus* – potential
357 role of host size. Ecology 66, 904–913.
- 358 Martinou, A.F., Seraphides, N., Stavrinides, M.C., 2014. Lethal and behavioral effects of pesticides on the
359 insect predator *Macrolophus pygmaeus*. Chemosphere 96, 167–173.

- 360 Monzò, C., Qureshi, J.A., Stansly, P.A., 2014. Insecticide sprays, natural enemy assemblages and predation on
361 Asian citrus psyllid, *Diaphorina citri* (Hemiptera:Psyllidae). Bull. Entomol. Res. 104, 576–585.
- 362 Moreno, D.S., Luck, R.F., 1992. Augmentative releases of *Aphytis melinus* (Hymenoptera: Aphelinidae) to
363 suppress California red scale (Homoptera:Diaspididae) in Southern California lemon orchards. J. Econ.
364 Entomol. 85, 1112–1119.
- 365 Najar-Rodriguez, A.J., Walter, G.H., Mensah, R.K., 2007. The efficacy of a petroleum spray oil against *Aphis*
366 *gossypii* glover on cotton. Part 2: indirect effects of oil deposits. Pest Manage. Sci. 63, 596–607.
- 367 Pan, H., Liu, Y., Liu, B., Lu, Y., Xu, X., Qian, X., Wu, K., Desneux, N., 2014. Lethal and sublethal effects of
368 cycloxaprid, a novel cis-nitromethylene neonicotinoid insecticide, on the mirid bug *Apolygus lucorum*. J. Pest
369 Sci. 87, 731–738.
- 370 Planes, L., Catalán, J., Tena, A., Porcuna, J.L., Jacas, J.A., Izquierdo, J., Urbaneja, A., 2013. Lethal and sublethal
371 effects of spirotetramat on the mealybug destroyer, *Cryptolaemus montrouzieri*. J. Pest Sci. 86, 321–327.
- 372 Prabhaker, N., Morse, J.G., Castle, S.J., Naranjo, S.E., Henneberry, T.J., Toscano, N.C., 2007. Toxicity of seven
373 foliar insecticides to four insect parasitoids attacking citrus and cotton pests. J. Econ. Entomol. 100, 1053–
374 1061.
- 375 Raciti, E., Saraceno, F., Siscaro, G., 2003. Mass rearing of *Aphytis melinus* for biological control of *Aonidiella*
376 *aurantii* in Sicily. IOBC/WPRS Bull. 26 (6), 125–134.
- 377 Retnakaran, A., Granett, J., Ennis, T., 1985. Insects growth regulators. In: Kerkut, G.A.,
378 Gilbert, L.I. (Eds.), Comprehensive Insect Physiology, Biochemistry and Pharmacology, vol. 12. Pergamon
379 Press, New York, pp. 529–601.
- 380 Rill, S.M., Grafton-Cardwell, E.E., Morse, J.G., 2008. Effects of two insect growth regulators and a
381 neonicotinoid on various life stages of *Aphytis melinus* (Hymenoptera: Aphelinidae). BioControl 53, 579–
382 587.
- 383 Rosen, D., DeBach, P., 1979. Species of *Aphytis* of the World (Hymenoptera:Aphelinidae), Shimitschek &
384 Spencer Ed., vol. 17, pp. 801.
- 385 Rosenheim, J.A., Hoy, M.A., 1986. Intraspecific variation in levels of pesticide resistance in field populations
386 of a parasitoid, *Aphytis melinus* (Hymenoptera:Aphelinidae): the role of past selection pressures. J. Econ.
387 Entomol. 79, 1161–1173.
- 388 Rosenheim, J.A., Hoy, M.A., 1988. Genetic improvement of a parasitoid biological control agent: Artificial
389 selection for insecticide resistance in *Aphytis melinus* (Hymenoptera: Aphelinidae). J. Econ. Entomol. 82,
390 1539–1550.
- 391 Saber, M., Abedi, Z., 2013. Effects of methoxyfenozide and pyridalyl on the larval ectoparasitoid *Habrobracon*
392 *hebetor*. J. Pest Sci. 86, 685–693.
- 393 Schneider, M., Smagghe, G., Pineda, S., Viñuela, E., 2004. Action of insect growth regulator insecticides and
394 spinosad on life history parameters and absorption in third-instar larvae of the endoparasitoid *Hyposoter*
395 *didymator*. Biol. Control 31, 189–198.
- 396 Siscaro, G., Longo, S., Lizzio, S., 1999. Ruolo degli entomofagi di *Aonidiella aurantii* (Maskell) (Homoptera,
397 Diaspididae) in agrumeti siciliani. Phytophaga IX, 41–52.
- 398 Sorribas, J., Garcia-Marí, F., 2010. Comparative efficacy of different combinations of natural enemies for the
399 biological control of California red scale in citrus groves. Biol. Control 55, 42–48.

400 Staal, G.B., 1975. Insects growth regulators with juvenile hormone activity. *Annu. Rev. Entomol.* 20, 417–460.

401 Sterk, G., Hassan, S.A., Baillod, M., Bakker, F., Bigler, F., Blumel, S., Bogenschutz, H., Boller, E., Bromand, B.,
 402 Brun, J., Calis, J.N.M., Coremans-Pelseener, J., Duso, C., Garrido, A., Grove, A., Heimbach, U., Hokkanen, H.,
 403 Jacas, J., Lewis, G., Moreth, L., Polgar, L., Roversti, L., Samsøe-Petersen, L., Sauphanor, B., Schaub, L., Staubli,
 404 A., Tuset, J.J., Vainio, A., Van De Veire, M., Viggiani, G., Vinuela, Vogt, H., 1999. Results of the seventh joint
 405 pesticide testing programme carried out by the IOBC/WPRS-working group “pesticides and beneficial
 406 organisms”. *BioControl* 44, 99–117.

407 Suma, P., Zappalà, L., Mazzeo, G., Siscaro, G., 2009. Lethal and sub-lethal effects of insecticides on natural
 408 enemies of citrus scale pests. *BioControl* 54, 651–661.

409 Tomé, H.V.V., Cordeiro, E.M.G., Rosado, J.F., Guedes, R.N.C., 2012. Egg exposure to pyriproxyfen in the
 410 tomato leaf miner *Tuta absoluta*: ovicidal activity or behavioural-modulated hatching mortality? *Ann. Appl.*
 411 *Biol.* 160, 35–42.

412 Vacas, S., Vanaclocha, P., Alfaro, C., Primo, J., Verdú, M.J., Urbaneja, A., NavarroLlopis, V., 2012. Mating
 413 disruption for the control of *Aonidiella aurantii* Maskell (Hemiptera: Diaspididae) may contribute to
 414 increased effectiveness of natural enemies. *Pest Manage. Sci.* 68, 142–148.

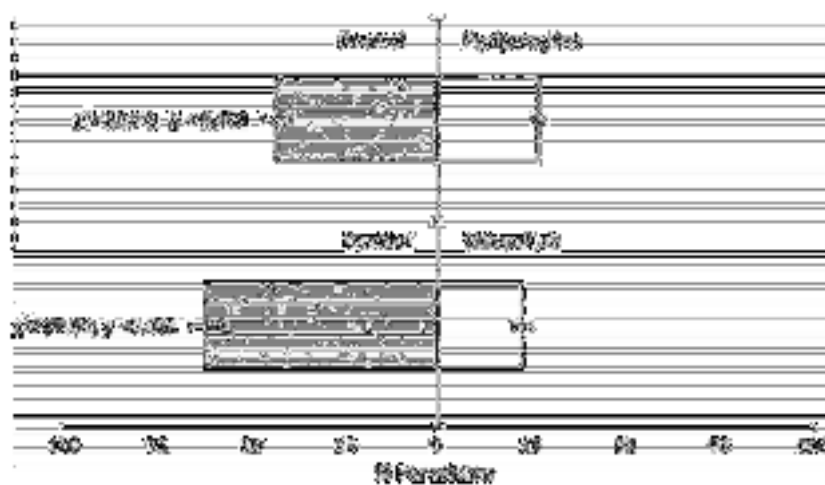
415 Vanaclocha, P., Papacek, D., Monzó, C., Verdú, M.J., Urbaneja, A., 2013a. Intra-guild interactions between
 416 the parasitoid *Aphytis lingnanensis* and the predator *Chilocorus circumdatus*: implications for the biological
 417 control of armoured scales. *Biol. Control* 65, 169–175.

418 Vanaclocha, P., Vidal-Quist, C., Oheix, S., Montón, H., Planes, L., Catalán, J., Tena, A., Verdú, M.J., Urbaneja,
 419 A., 2013b. Acute toxicity in laboratory tests of fresh and aged residues of pesticides used in citrus on the
 420 parasitoid *Aphytis melinus*. *J. Pest Sci.* 86, 329–336.

421 Vasquez, C.J., Morse, J.G., 2012. Fitness components of *Aphytis melinus* (Hymenoptera: Aphelinidae) reared
 422 in five California insectaries. *Environ. Entomol.* 41, 51–58.

423 Zappalà, L., 2010. Citrus integrated pest management in Italy. In: Ciancio, A., Mukerji, K.G. (Eds.), *Integrated*
 424 *Management of Arthropod Pests and Insect Borne Diseases*. Springer, pp. 69–96.

425 Zappalà, L., Campolo, O., Grande, S., Saraceno, F., Biondi, A., Siscaro, G., Palmeri, V., 2012. Dispersal of *Aphytis*
 426 *melinus* (Hymenoptera: Aphelinidae) after augmentative releases in citrus orchards. *Eur. J. Entomol.* 109,
 427 561–568.



428
 429 Fig. 1. Mean parasitism rates (\pm SE) in the dual-choice test between treated and untreated hosts by *Aphytis*
 430 *melinus*.

Table 1

Mean (\pm SE) of the lethal and sublethal toxicity on reproduction of mineral oil and pyriproxyfen on *Aplysia melinae* adults and young instars. Within rows, data followed by the same letter are not significantly different ($P < 0.05$; LSD test).

Trial	Exposed instar	Mineral oil	Pyriproxyfen	Control
Acute toxicity (% mortality)	Adults	97.2 \pm 1.0 a	36.4 \pm 5.6 b	11.4 \pm 4.4 c
	Young instars	13.7 \pm 3.6 a	17.0 \pm 6.1 b	4.7 \pm 5.1 a
Sublethal effects on reproduction (# of progeny/female/week)	Adults	-	42.8 \pm 11.1 a	48.5 \pm 10.1 a
	Young instars	10.9 \pm 5.4a	14.2 \pm 6.1 b	31.1 \pm 5.9 a

Table 2

Mean (\pm SE) of sex ratio (# females/total progeny) of (i) the progeny of *Aplysia melinae* adults survived to the residual exposure to pyriproxyfen, (ii) adults developed from larvae exposed to mineral oil and pyriproxyfen and (iii) of their progeny. Within rows, data followed by the same letter are not significantly different ($P < 0.05$; LSD test).

Trial		Mineral oil	Pyriproxyfen	Control
Progeny of adults surviving the exposure		-	0.55 \pm 0.07 a	0.69 \pm 0.04 a
Young instars exposure	Emerging adults	0.64 \pm 0.03 a	0.67 \pm 0.07 a	0.61 \pm 0.03 a
	Progeny	0.65 \pm 0.03 a	0.46 \pm 0.09 b	0.70 \pm 0.04 a