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1 **A Comprehensive Overview on Microalgal-Fortified/Based Food and Beverages**

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32 **ABSTRACT**

33 Microalgae produce a variety of compounds that can be used for aquaculture, nutraceutical
34 purposes, and third-generation biofuel sectors. Moreover, they have been considered for the
35 production of fortified food and beverages claiming to be healthier than other foods. Using
36 microalgal oil or incorporating microalgae biomass or their metabolites in food and beverages
37 provides health benefits due to properties such as anti-inflammatory, antioxidant, immune-
38 enhancing, and to their role against various diseases such as cardiovascular metabolic,
39 atherosclerosis, and hypertension. This review focuses on the worldwide research carried out about
40 the incorporation of microalgae – either biomass or their high-value compounds – in food and
41 beverages, and on the microalgal fortified/based food and beverages currently present worldwide on
42 the market. The metabolites bioavailability aspect and the current legislation are considered. There
43 is an ever-growing interest in this field but the volume of production is still limited. Various
44 challenges, one of which is the cost of producing biomass, need to be overcome for a profitable
45 market. More effort, involving different expertise, is needed to improve the microalgal production
46 system, from cultivation to harvesting and biorefinery, to produce improved novel products.

47

48 **KEYWORDS** Microalgae; biomass; metabolites; food, beverages; fortification; bioavailability;
49 legislation

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56 **Introduction**

57 Microalgae are widespread, photosynthetic microorganisms able to grow rapidly using, similarly to
58 plants, light, sugars, CO₂, N, P, and K. Some of them can also grow heterotrophically (organic
59 carbon sources are used for metabolism) and mixotrophically (inorganic and organic carbon
60 sources are concurrently used for metabolism). They include algae *sensu stricto* and
61 cyanobacteria.[1] The former are eukaryotes and the most abundant are the diatoms
62 (Bacillariophyceae), the green algae (Chlorophyceae), and the golden algae (Chrysophyceae). The
63 latter – Cyanophyceae or blue-green algae – are prokaryotes and, in this review, they are referred to
64 as microalgae. There are thousands of different species of microalgae and cyanobacteria. In
65 particular, it has reported the existence of 50,000 species of which 30,000 have been studied and
66 analyzed.[2] Diatoms dominate in phytoplankton life and with more than 100,000 species are
67 considered the largest group of biomass producers.[3] Among the thousands of species of
68 microalgae that are believed to exist, only few species are industrially cultivated to produce useful
69 chemical compounds[3,4] such as lipids, proteins, carbohydrates, vitamins, and pigments as well.

70 Microalgae have been considered for nutraceutical compounds production[5,6] and also for
71 third-generation biofuels[7] as both in monoculture[8,9] and in mixed culture cultivation.[10–12]
72 Microalgal high-value biological active compounds – pigments, antioxidants, β -carotenes,
73 polysaccharides, triglycerides, and vitamins – have been reviewed[13] and they are used in various
74 sectors such as the food production.[2,14–16] Enzing et al.[3] reported three main ways to use
75 microalgae in the food sector: whole dried microalgae as dietary supplement, high-value molecules
76 extracted from microalgae, and dried microalgae biomass added to new food formulations. The use
77 of microalgal biomass for human consumption is allowed for few species and it is related to food
78 safety regulations.[14]

79 Microalgae commonly cultivated as additives for human foods are mainly chlorophyta and
80 cyanobacteria.[17–19] In particular, *Chlorella* sp., *Haematococcus pluvialis*, *Dunaliella salina*,

81 *Aphanizomenon flos-aquae*, *Arthrospira* sp. (formerly *Spirulina*), *Schizochytrium* sp.,[20,21] and
82 *Scenedesmus* sp.[22] are used for human nutrition.

83 Proteins produced by microalgae have high nutritional value because of their composition of
84 essential amino acids.[17,23] High content of proteins is reported for *Chlorella* (up to 58% of dry
85 weight)[17] and *Arthrospira* (50–70% of dry weight)[24] (Tables 1–3).

86 In the food industry sector, polysaccharides are mainly derived from macroalgae. An
87 exopolysaccharide produced by the microalga *Porphyridium cruentum* can substitute for
88 carrageenans.[17]

89 Polyunsaturated fatty acids (PUFAs) such as γ -linolenic acid (GLA) and long chain PUFAs
90 such as eicosapentaenoic (EPA), docosahexaenoic (DHA), and arachidonic (AA) acids have
91 important role in human health, giving protection against cardiovascular, coronary and heart
92 diseases, atherosclerosis, metabolic diseases, rheumatism, skin diseases, and cancer.[22,25,26]
93 *Chlorella minutissima* produces high content (45% w/w) of EPA.[27] *Arthrospira* is a good source
94 of GLA (8–31.7%).[28] Hoseini et al.[24] have reported values of 10–20% and 49% of GLA for
95 *Arthrospira maxima* and *Arthrospira platensis*, respectively. *Nannochloropsis* sp., *Isochrysis* sp.,
96 *Nitzschia* sp., *Phaeodactylum* sp., and *P. cruentum* (up to 27% of fatty acids) produce EPA (Table
97 1), *Porphyridium* sp. produce AA (up to 36% of fatty acids), while *Cryptocodinium* sp. (40–50%
98 of fatty acids), *Ulkenia* sp., and *Schizochytrium* sp. produce DHA (Table 1).[5,14,20,29–32]
99 *Pavlova lutheri* produces EPA and DHA (Table 1).[33] Patents for producing DHA by
100 *Cryptocodinium cohnii* are reported.[34] Also, other PUFAs than DHA from microalgae are used
101 in the production of infant formula and functional food.

102 Microalgae also produce chlorophylls, carotenoids (β -carotene, astaxanthin, lutein, and
103 zeaxanthin), and phycobiliproteins (phycocyanin, phycoerythrin). These compounds are pigments
104 that can be used both as food colorants, thus replacing various synthetic dyes, and as food

105 supplement for important properties such as anti-inflammatory, antioxidant, hypolipidemic,
106 enhancing the immune system function, and inhibiting tumor progression.[35]

107 Astaxanthin is produced by *H. pluvialis* (up to 0.2–2.0% on dry weight basis)[20,36–38] and by
108 *Euglena cf. sanguinea* (80% of the carotenoid pool),[18] lutein by *Muriellopsis* sp. (4–6
109 mg/g),[39,40] and β -carotene is produced by *D. salina* (Table 2).[20,22,38,41] Pangestuti and
110 Kim[42] have reported phycobiliproteins from marine algae; phycocyanin is mainly produced by *A.*
111 *platensis* (Table 2)[24,43,44] and *Synechococcus* sp. (80–100 mg/g)[45] while phycoerythrin by *P.*
112 *cruentum* (32.7% dry weight)[46] and *Rhodospirillum rubrum* (8.0% dry weight).[47] Phycocyanin is
113 mainly reported to be used as food colorant and as supplement for health food.[48]

114 Moreover, microalgae – first *Arthrospira* sp. and *Chlorella* sp. among others – are a source of
115 vitamins such as A, B1, B2, B6, B12, C, E, folic acid, and pantothenic acid (Table 3).[14,49]
116 Vitamins produced in more concentration are B3 (range 7.8–23.8 mg/100 g dry weight for
117 *Isochrysis galbana* and *Chlorella*, respectively), C (range 10.0–19.1 mg/100 g dry weight for
118 *Chlorella stigmatophora* and *Tetraselmis suecica*, respectively), and E (range 0.00045–66.9 mg/100
119 g dry weight for *Schizochytrium* sp. and *C. stigmatophora*, respectively).

120 Nowadays, there is an ever-growing interest in the application of microalgae for the
121 development of healthier, innovative, and attractive food products. Nevertheless, the production and
122 distribution of new formulates are still limited[50] even if since the year 2000, the microalgae
123 cultivation for food and feed production had a fivefold increase.[51]

124 Important aspects to consider in the development of novel microalgal-based food/beverage
125 formulations are the cost-effectiveness of the sector, the biological availability of functional
126 compounds from microalgae and their stability throughout the life of the products,[52] and the
127 consumers' perception and acceptance of the microalgal-based products. This paper aims to
128 contribute, as an overview, to the knowledge on the sector of novel food/beverage formulates from

129 microalgae as (1) biomass incorporated to foods and (2) metabolites extracted from microalgae and
130 incorporated in foods. Moreover, challenges to face up in order to improve this novel sector are
131 discussed.

132

133 **Applications to the food sector**

134 At present, the technology of production, recovery, and transformation of microalgae utilizes low
135 volumes with associated high cost. For this reason, the extraction of high-value compounds to be
136 used as supplements and nutraceuticals for human nutrition is considered the most encouraging
137 sector compared to the bulk production of carbohydrates and proteins, which needs higher volume
138 of production and, therefore, cost-effective scale-up process in order to reduce the costs. Moreover,
139 the high-value compounds from microalgae are more effective for food applications compared to
140 the synthetic alternatives. This is determined by their chemical conformation, which makes them
141 more effective for specific applications, such as in infant formula and dietary supplements.[20]
142 There are also many examples of the use of whole microalgae biomass incorporated in food, and
143 this confirms the active industry research in order to reduce the cost and, at the same time, increase
144 the volume of production. The microalgal concentration of valuable compounds (Tables 1–
145 3),[13,20,53–79] often higher compared to other sources, can influence positively human health and
146 express the real potential use of microalgae in the food sector.

147 The regulation about the use of microalgae as food/supplement varies among countries (Table
148 4). In the European Union (EU), until now, novel foods (food not consumed to a significant degree
149 in the EU prior to 15 May 1997) and food ingredients are regulated by the Regulation (EC) No.
150 258/97.[80] *Chlorella pyrenoidosa*, *Chlorella vulgaris*, and *Heterochlorella luteoviridis*, having
151 been on the market and consumed before 15 May 1997, are not subject to the Novel Food
152 Regulation.[81] Under Regulation (EC) No. 258/97, the oil from *Schizochytrium* sp. rich in DHA
153 and EPA has been allowed to be placed on the market in Europe as a novel food ingredient.[82–84]

154 Also, astaxanthin from *H. pluvialis* was approved as novel food ingredients in 2006, *Tetraselmis*
155 *chuii* was approved as novel food to be used as sauce, special salt, and condiment under Regulation
156 No. 258/97 in 2004, and the oil from *Ulkenia* sp. was approved in 2009 as a novel food ingredient
157 in bakery products, cereal bars, and nonalcoholic beverages including milk-based beverages.[85]
158 Current applications for food/food ingredient concern the microalga *Nannochloropsis gaditana* and
159 the EPA-rich oil derived from the microalga *Phaeodactylum tricornutum*. In order to simplify the
160 current authorization procedures taking into account recent developments in Union law and
161 technological progress, a new regulation on novel food was established with effective validity at the
162 end of 2017.[86] According to this regulation, a novel food falls into different categories among
163 which is a “food consisting of, isolated from or produced from microorganisms, fungi or algae.”

164 In the United States, the Food and Drug Administration (FDA) classified some microalgae as
165 generally recognized as safe.[87–94] Examples are *Euglena gracilis* (for conventional food and
166 beverage products such as baked, cereal, dairy and milky products, processed fruits, fruit juices, soft
167 candy, and soups), *Arthrospira* sp. (for breakfast/power/energy bars, smoothies, and condiment for
168 salads and pasta), DHA oil from *Schizochytrium* sp. (for soy protein bars, processed vegetable
169 drinks, hard and soft candy, nondairy and powdered cream substitutes, jams and jellies, milk and
170 flavored milk, and soy milk), DHA oil from *Ulkenia* sp. (for addition in a vast variety of foods), *D.*
171 *salina* powder (for bread, rolls, cookies, crackers, cheese, mayonnaise, tofu, and fermented soybean
172 products), *Auxenochlorella protothecoides* (formerly *Chlorella*) (algal flour for baked goods,
173 beverages and beverage bases, dairy product analogs, egg products, fats and oils, sauces, processed
174 vegetables, vegetable juices, and soups), and *C. vulgaris* (for power bars, sport beverages, energy
175 drinks, and low fat soy milk).

176 In Australia, the Food Standards Australia New Zealand approved DHA-rich oil from
177 *Schizochytrium* sp. and *Ulkenia* sp. as novel food ingredients.[95]

178 In Japan, the Ministry of Health, Labour and Welfare defined the specifications and standards
179 for food additives where are reported the approved food additives.[88] *Chlorella* powder is
180 generally provided as food and food additives – color, flavor, and taste enhancer. Also, in the list of
181 food additives appear the phycocyanin from *Arthrospira* sp., carotene from *Dunaliella* sp., and the
182 microalga *Haematococcus* as colorant.

183 Microalgae can be used both as biomass and, after extraction processes from the biomass, as
184 pigments and oils in a variety of food and beverages (Figure 1). Applications of microalgae in
185 different food and beverage categories are considered below.

186

187 **Microalgae added to dairy and probiotic products**

188 The incorporation of microalgal biomass to these products aims to increase their nutritional
189 properties also via the stimulation of the lactic microflora typical of these products. Cheeses and
190 cheese analogs, also called processed/imitation cheeses, are products in which the milk components
191 are partially or wholly replaced by non-milk-based components, improved for their functional role
192 by the addition of dried microalgal biomass.[96] Positive effects of the addition of *Chlorella*
193 biomass (0–2%) into Appellenzer cheese have been reported on lactic acid bacteria (LAB) viability:
194 a biomass concentration of 0.5% positively influences the quality and sensory properties.[97] The
195 use of *Arthrospira* – also decolorized, odorless, and tasteless biomass – is used to produce cheese,
196 tofu and yoghurt.[98] The incorporation of *Chlorella* biomass (0.5%) during cheese production
197 improved physical–sensory properties.[99,100] Recently, Shalaby and Yasin[101] reported the
198 formulation of innovative croissant stuffed with cheese obtained with *Chlorella* spp.; the 2–3% of
199 the microalgae improved croissants taste, texture, and quality.

200 Microalgae, especially *C. vulgaris* and *A. platensis*, in coculture with LAB are used in yoghurt
201 and probiotic fermented milks production. Various authors have reported the stimulation role that

202 microalgae have on the growth of probiotic bacteria both in the products and in the human
203 gut.[102–104] Stimulation of the growth of LAB in fermented milk incorporated with microalgae
204 has been reported.[16,105–108] Adding *C. vulgaris* and *A. platensis* at different concentrations to
205 yoghurt determined higher viability of two probiotic LAB strains of *Lactobacillus acidophilus* LA-
206 5 and *Bifidobacterium lactis* BB-12 than the product free of microalgae both at the end of
207 fermentation and throughout the storage.[109] On the other hand, the same authors suggest that
208 using high concentration of microalgae confers unpleasant color and graininess to the product.
209 Therefore, using low concentration of microalgae gives better sensory characteristics.
210 Supplementing fermented milk with microalgae could give a way to sensory defects that can be
211 avoid by adding flavors, texture improvers, and colors.[110] Varga et al.[111] have reported that *A.*
212 *platensis* added to fermented milk has positive effect on *Bifidobacterium* sp. showing also
213 antifungal effect on spoilage yeasts and molds. Novel yoghurt products, frozen with low fat, high
214 protein, supplemented with *Arthrospira*, and papaya pulp, have been reported.[112] In particular,
215 the best sensory characteristics were scored by the formulation with 6% of *Arthrospira* and 10% of
216 papaya. Functional drinkable skimmed milk yoghurts added with *Chlorella* have been
217 reported.[113] The authors tested both microalgal powder (0.25%) and microalgal liquid extract
218 (2.5–10%); in both cases, the storage of the yoghurt at 4°C maintained the quality in terms of pH,
219 titratable acidity, and viable count of LAB. Even so, the yoghurt with no addition showed higher
220 sensory acceptability than the other formulates. Recently, authors reviewed the use of *C. vulgaris*
221 and *A. platensis* to produce functional dairy products.[110,114] One of the requirements of
222 probiotic fermented milks is the high number of viable LAB; therefore, any supplementation must
223 positively affect this aspect other than the sensory properties of the final product. The microalgae
224 were added in the range of 0.25–1%; generally, this supplementation increased probiotic bacteria
225 viability but only the product with low concentration of microalgal biomass gained good sensory
226 attributes. Due to this, despite having health benefits, consumers may not accept the product due to
227 its strange color and its graininess. Possible solutions could be the addition of colorants to

228 homogenize the discolorations and the addition of texture improvers to solve the off-texture
229 problems. In addition, the incorporation of microalgae determines a price increase of the final
230 product; so, the possible solution is a more costeffective production of the microalgae.

231 The effective release of the compounds from cells and how it is affected by the food production
232 and processing techniques are aspects to consider for compound availability both for human and
233 lactic microflora.

234

235 **Microalgae added to bakeries, pasta, dressing, and beverages**

236 Growing interest in the market for new food products with microalgae and the variety of foods that
237 can be fortified with microalgae – pasta, biscuits, bread, noodles, snack foods, candy or gums, drink
238 mixes, soft drinks, soups, and sauces – has motivated other researches in the field.[49,98]

239 The development of soups, noodles, bread and rolls, biscuits, ice cream, soy sauce, and
240 powdered green tea based on *Chloroidium ellipsoideum* (formerly *Chlorella ellipsoidea*) has been
241 reported.[2] The addition of *A. platensis* (10%) to conventional bread enhanced the nutritional
242 quality and the shelf life of the bread without negative impact on sensory evaluation. In particular,
243 the microalgal-enriched bread resulted in higher proteins and volatile compounds content. The
244 sensory assessment was satisfactory even perceiving algal taste.[115] Adding *Chlorella* powder to
245 white bread determined an improvement of its quality characteristics in terms of volume and color
246 while the texture was not significantly affected. The bread with 0.2% of *Chlorella* powder scored
247 high sensory evaluation.[116] Novel cookies and pasta enriched with *Nannochloropsis oculata* were
248 characterized by good concentration of PUFAs (e.g., cookies enriched with 3% of biomass had a
249 value of DHA + EPA of 298 mg/100 g). Regarding the sensory evaluation, biomass incorporation

250 up to 2% and 3% was positively evaluated for cookies and pasta, respectively.[117] Bakery
251 *Arthrospira* products with 2% of microalga such as rice pasta, sfogliatine, and grissini are produced
252 in Italy (Microlife Nutrition).

253 Novel biscuits rich in functional compounds were developed with the addition of *C.*
254 *vulgaris*,[118,119] *H. pluvialis*, and *I. galbana*. [17,120] Industrial production of traditional Iranian
255 biscuits supplemented with *A. platensis* was reported; the biscuits fortified with 1.0–1.5% of
256 microalga exhibited improved and desirable functional and sensory characteristics.[121]
257 *Arthrospira* powder was also used as ingredient for an orange-flavored chewable wafer[98] and used
258 by the industry Microlife Nutrition (Italy) to produce chocolate bars and biscuits (Table 5).
259 *Arthrospira* sp. was used at the concentration of 2.6% to produce snacks with improved nutritional
260 content (increase of 22.6% in proteins, 28.1% in lipids, and 46.4% in minerals) without
261 significantly affecting physical parameters and the sensory quality with an acceptability index of
262 82%. Also, these snacks showed physical and microbiological stability during one year of
263 storage.[122] *A. platensis* as extract or as phycocyanin can be incorporated in sour milk and green
264 tea.[123] Kim and Chung[124] have evaluated the addition of *Chlorella* powder (0–4%) to make a
265 yellow layer cake; the novel cake exhibited improved quality from textural, sensory, and functional
266 point of view.

267 *A. protothecoides* is the component of AlgaVia® Lipid-Rich Whole Algae (algal flour) and
268 AlgaVia® Protein-Rich Whole Algae (algal protein) (Corbion, The Netherlands). These food
269 ingredients deliver lipids rich in polyunsaturated fats, proteins plus fibers, and micronutrients,
270 respectively, and they are used to make various types of food (Table 5). AlgaVia® Lipid-Rich
271 Whole Algae is commercially available throughout North America (US, Canada, Mexico), the EU,
272 the United Kingdom, Australia/New Zealand, and Brazil while AlgaVia® Protein-Rich Whole
273 Algae is not commercially available in Brazil.

274 Fradique et al.[125] have reported the enrichment of fresh spaghetti with biomass of *C. vulgaris*
275 and *A. maxima*. The supplementation (0.5–2%) with these microalgae resulted in an improvement
276 of cooking and textural properties; the addition of microalgae resulted in an increase in the raw
277 pasta firmness and its color remained relatively stable after cooking. These microalgae pastas had
278 higher acceptance scores for colors, similar to pastas produced with vegetables, and good cooking
279 and textural properties.

280 Different Italian companies produce pasta with *Arthrospira* (Table 5) such as tagliolini, rice
281 fusilli, corn penne (2% of microalga) (Microlife Nutrition), corn flour conchiglie (10% of
282 microalga) (Nutracentis), wheat fusilli, kamut penne, and spelt spaghetti (3% of microalga) (La
283 finestra sul cielo). In addition, French companies produce novel pasta such as tagliatelle duo curry
284 and Spirulina (Cornand) and Tagliatelles with *Spirulina* (GlobeXplore) (Table 5).

285 A total of 328 new packaged food, drink, and pet food products with *Chlorella* were launched
286 recently in Japan, United States, South Korea, Canada, Germany, and other countries.[126]

287 The incorporation of microalgal biomass is an interesting and promising alternative to the
288 production of conventional oil-in-water emulsions such as mayonnaises and salad dressings. Main
289 advantages of the microalgal use are natural color, source of biologically active compounds, and
290 antioxidant effect. The possibility to use microalgal biomass as coloring agent in oil-in-water
291 emulsions has been reported.[127] In these kinds of food, the trend is to replace the egg yolk
292 lipoproteins with pea proteins in order to reduce the fat content. Studies on the interaction between
293 pea proteins and microalgal biomass have been carried out to conclude that the presence of the
294 biomass does not destabilize the product texture; on the contrary, it can present a synergistic effect
295 improving the texture and sensory properties.[128] Green and orange *C. vulgaris* biomass and *H.*
296 *pluvialis* biomass are reported to be suitable for the oil-in-water vegetable food emulsions since
297 they give stable and attractive color, good texture, and antioxidant effect.[129] *A. maxima* and
298 *Diacronema vlkianum* were used to produce novel vegetable gel desserts rich in PUFAs and

299 improved for texture.[123] Batista et al.[130] have studied the effect of the addition of *H. pluvialis*
300 and *A. maxima* on the viscoelastic behavior of gel vegetarian food in order to set the best conditions
301 for these kinds of foods.

302 *Chlorella* powder was used to improve the quality of some traditional Korean foods: Mul-
303 kimchi, a drinkable broth made with fermented vegetables, added with 0.05% of *Chlorella*
304 exhibited increased LAB population, good sensory characteristics, and improved shelf life during
305 refrigerated storage[131]; Sulgidduk, a steamed rice cake, supplemented with 0.2% and 0.5% of
306 *Chlorella* exhibited the best characteristics of quality.[132]

307 Microalgae of Chlorophyceae family including *Chlorella*, *Scenedesmus*, and *Chlorococcum*
308 can be used together with *Saccharomyces cerevisiae* to make an alcoholic beverage with pleasant
309 and wine-like taste.[133]

310 Microalgal biomass incorporation is an appealing alternative to the conventional production.
311 Generally, this fortification is accepted if the improvement of the healthiness characteristics
312 corresponds to an equal or improved level of sensory characteristics compared to the conventional
313 products. According to the product considered, the expectations are different (the absence of
314 graininess for yoghurt, homogeneity in texture for pasta and mayonnaises); therefore, the viability
315 of these novel food and beverage products depends on a balance between nutritional and sensory
316 requirements.

317 Different from the previous sections, reporting the incorporation of microalgal biomass on
318 food, the two following sections report the use of microalgal pigments and oils extracted from the
319 biomass.

320

321 **Pigments from microalgae added to miscellaneous products**

322 One of the characteristics of microalgae is their color due to the presence of various types of
323 pigments. Other than chlorophylls, which are the primary photosynthetic pigments, they can have
324 carotenoids and phycobiliproteins (Figure 2). These natural pigments are important in the food
325 sector since they can be used both to give color and for their antioxidant properties.

326 Among microalgal pigments, phycocyanin, mainly produced by *A. platensis*, is used in the food
327 industry especially in dairy and milky products (fermented milk, milk shakes, and ice creams),
328 jellies, coated soft candies, soft drinks, health drink, sour milk, green tea, confectionaries (blue
329 smarties), desserts, and sweet cake decorations.[16,44,134] The blue pigment phycocyanin of the
330 red microalga *Porphyridium aeruginum* is used mainly in drinks such as Pepsi® and Bacardi
331 Breezer®[44] due to its stability to pH changing and to light exposure (Table 5). Phycocyanin
332 derived from *Arthrospira* is used to produce Linablue® as natural blue food colorant (DIC
333 Corporation). This product can be used for beverages, frozen desserts, macarons, and candy-coated
334 chocolates (Table 5). Sekar and Chandramohan[44] have reported three patents concerning the use
335 of phycobiliproteins in drink/beverage compositions: CN 1127611,[135] CN 1096178,[136] and
336 WO 03099039.[137]

337 Table 5 reports the use of β -carotene and astaxanthin, their health and technological aspects,
338 and the producer companies.

339

340 **Oil from microalgae added to different food**

341 The main source of long chain PUFAs, especially ω -3, is fish oil; however, the use of this oil as
342 food additive gives problems since it confers fishy smell and taste and poor oxidative stability. A
343 good alternative is the use of oil from various species of microalgae. Also, even though microalgal
344 oil can undergo lipid oxidation, compared to fish oil, it is richer in vitamins and also in other
345 substances such as carotenoids and phenolics that can protect it from oxidation.[138]

346 Valencia et al.[139] have reported significant enrichment of ω -3 PUFAs in a new formulation
347 of dry fermented sausages made incorporating *Schizochytrium* sp. oil partially substituting pork
348 backfat. Healthier dry fermented sausage formulations were obtained by the incorporation of mixed
349 linseed and algae oils.[140]

350 Oil emulsion from microalgae was used to enrich strawberry flavored yoghurt,[141] ice
351 cream,[142] and milk.[143] The yoghurt and ice cream supplemented with algae oil emulsion to
352 provide 500 mg ω -3 fatty acids per 272 g serving and 300 mg ω -3 fatty acids per 65 g serving,
353 respectively, scored moderate acceptability by trained panel due to distinguishable strong fishy
354 flavor.

355 According to the Commission Implementing Decision (EU) 2015/545/CE and
356 2015/546/CE,[82,83] DHA and EPA-rich oil from *Schizochytrium* sp. can be used in dairy products
357 except milk-based drinks (200 mg/100 g or for cheese products 600 mg/100 g), dairy analogs except
358 drinks (200 mg/100 g or for analogs to cheese products 600 mg/100 g), spreadable fat and dressings
359 (600 mg/100 g), breakfast cereals (500 mg/100 g), food supplements (250 or 450 mg DHA per day
360 for normal population and pregnant women, respectively), foods for energy restricted diets for
361 weight reduction (250 mg per meal replacement), and other foods for particular nutritional uses
362 (200 mg/100 g), bakery products (breads and rolls), and sweet biscuits (200 mg/100 g), cereal bars
363 (500mg/100 g), cooking fats (360 mg/100 g), non-alcoholic beverages (80 mg/100 ml), infant
364 formula and follow-on formula (in accordance with Directive 2006/141/EC), processed cereal-
365 based foods, and baby foods for infants and young children (200 mg/100 g). The DHA-rich
366 microalgae oils from *Schizochytrium* sp. and *Ulkenia* sp. can be used as food ingredients in a variety
367 of foods and in infant formula.[144,145]

368 *C. cohnii* cells containing DHA-rich oil can be used in baby foods and infant formula; many
369 of the latter containing Martek Corporation's (USA) DHA-rich oil are available worldwide –United
370 States, United Kingdom, Mexico, China, and Canada[32] (Table 5). A microalga, originally found

371 in the sap of a chestnut tree in Germany, is cultivated and used by the company Corbion (The
372 Netherlands) to extract oil to produce the AlgaeWise® Ultra Omega-9 Algae Oil that can be used as
373 culinary oil – also for frying – and spray coatings and to make margarines and spreads, sauces and
374 dressings, and bakery. This microalga is also used to produce the AlgaeWise® Algae Butter
375 structuring hard-stock (Corbion) whose application is in the bakery industry, spreads, and
376 confectionery (Table 5).

377 To affirm effectively the usefulness of microalgal fortified products, knowledge about the fate
378 of the microalgal compounds after the food ingestion is fundamental. The following section gives
379 the state-of-art of the microalgae and their valuable compounds bioavailability.

380

381 **Bioavailability of functional compounds from microalgae**

382 The term bioavailability was defined by the FDA as “the rate and extent to which the active
383 ingredient is absorbed and becomes available at the site of action.”[146] After food ingestion, the
384 bioavailability involves the gastrointestinal digestibility and solubility of the food elements, the
385 adsorption by intestinal endothelium, the distribution by circulatory system, and the utilization by
386 the target site. Also, bioavailability is divided as bioaccessibility, which examines compounds of a
387 whole food released into the gastrointestinal tract to identify those elements effectively accessible
388 for adsorption, and bioactivity, which refers to the intestinal adsorption of food elements, the
389 transport to the target site, the elements biotransformation at the target site, and the physiological
390 response.[147] Bioavailability, bioaccessibility, and bioactivity are assessed by different *in vitro*
391 and *in vivo* studies that need to be harmonized to obtain univocal data. Considering protein
392 digestibility, for example, a harmonizing attempt is the INFOGEST, a static *in vitro* digestion
393 model.[148,149] Moreover, the data on bioavailability often refer to seaweeds and algal
394 supplements while the knowledge about the bioavailability of compounds from microalgae, also as
395 biomass incorporated into a food matrix, is still limited. Important aspects to consider in view of

396 compounds release and availability, the real assimilation during the digestion process and their
397 interaction with the gut microbiome,[52] are the effect of the food processing techniques on the
398 algal compounds and the interaction among them and the food matrix.

399 Becker[150] reported that *Scenedesmus obliquus*, *Spirulina* sp., and *Chlorella* sp., exhibited
400 protein digestibility coefficient values of 88.0%, 77.6%, and 76.6%, respectively, in comparison
401 with casein and eggs that have higher coefficients. The protein digestibility of *S. obliquus*, *C.*
402 *vulgaris*, *Nannochloropsis granulata* increased by disrupting cells with different methods[151–154]
403 or pH-shift process for *N. oculata*[155] while the protein digestibility of *A. platensis* was greater for
404 the fresh biomass compared to the treated one. Little differences in protein digestibility were
405 reported for *Nostoc commune* treated and untreated biomass.[156] Also, the presence in the
406 microalgal cell of high level of polysaccharides and polyphenols causes a reduction of protein
407 digestibility.[157,158]

408 Concerning the bioavailability of PUFAs, it is important to consider their presences in the
409 different classes of lipids (polar and neutral), their positions in the triacylglycerols,[159] and their
410 bonds with phospholipids or glycolipids.[160] In nutrient-starved cells of *Nannochloropsis*, the
411 neutral lipids (as storage molecules function) can contain 90% or more of triacylglycerols-rich in
412 C16 series of fatty acids, which have limited value as food ingredient. During starvation, the
413 mechanisms of degradation of polar lipids (as structural molecules function) and of inter-conversion
414 and repackaging of PUFAs such as EPA could have a role in distributing the ω -3 fatty acids in the
415 different classes of lipids with possible modification of bioavailability in food products.[161] The
416 position of fatty acids in the triacylglycerols affects the nutritional properties, the adsorption, and
417 the organism metabolism. In fact, the triacylglycerols with PUFAs in sn-2 position have more
418 nutritional properties and they are better adsorbed compared to triacylglycerols with PUFAs
419 casually rearranged.[159] It has been reported that PUFAs bound to phospholipids are more
420 bioavailable than those bound to triacylglycerols.[162] Kagan et al.[160] studied the appearance of

421 EPA and DHA in human plasma after the ingestion of breakfast followed by oil from *N. oculata*
422 (rich in EPA conjugated to phospholipids and glycolipids) and krill oil (rich in EPA and DHA
423 conjugated to phospholipids). The fatty acids appeared in plasma 3 h after breakfast and the
424 concentration of EPA was higher with the ingestion of microalga oil; this could be attributable to
425 the bond with glycolipids as a more effective system to deliver EPA to human. Using in vitro
426 digestion models, Cavonius et al.[155] have reported that intact cells of *Nannochloropsis* were not
427 accessible and that cell disruption coupled with pH-shift processing with protein solubilization
428 increased the accessibility of lipids. Meal supplementation with *D. vlkianum* (single-dose of 101
429 mg/kg EPA + DHA, human equivalent dose of 24.2 g of microalga) in rats increased the fatty acids
430 content in erythrocytes, liver, heart and adipose tissues, and in serum.[163] Clinical trials testing
431 DHA supplements from *C. cohnii* and *Schizochytrium* along with a snack bar fortified with
432 supplement from *Schizochytrium* highlighted that all DHA forms were equally bioavailable with a
433 dose–response relationship. In addition, the similar plasma DHA values between capsules and
434 fortified snack bars suggest that consuming DHA supplement with food increases its
435 bioavailability.[164] Another study providing algal-DHA oil with orange juice concluded that
436 potentially food may favor the DHA adsorption.[165] The type of food matrix appears to have a
437 role in the ω -3 bioavailability; in particular, nanoemulsion technology using yoghurt as a food
438 vehicle caused a rapid increase in DHA levels.[166]

439 Fabregas and Herrero[77] reported that the concentration of vitamins (E, B1, and β -carotene)
440 from *T. suecica*, *I. galbana*, *Dunaliella tertiolecta*, and *C. stigmatophora* exceeded the
441 concentration present in food recognized as source of these vitamins. For fat-soluble vitamins, it
442 was reported that the bioavailability increases consuming lipid-rich foodstuffs.[167] Also, the
443 vitamins bioavailability increases by microalgal biomass disruption.[168]

444 The efficiency of carotenoid absorption is affected by food matrix, type of processing, other
445 food components, and nutritional and physiological status.[169] The bioaccessibility of *N. oculata*

446 and *Chaetoceros calcitrans* was significantly higher in dried extract form than the powdered
447 form.[168] Gille et al.[170] reported different roles of biomass treatment on carotenoids
448 bioaccessibility from two microalgae species. *Chlamydomonas reinhardtii* is a good carotenoid
449 source without processing, while processing methods are necessary for *C. vulgaris*. β -carotene in
450 *Spirulina* is considered more digestible than the vitamin in leafy green vegetables given its
451 unicellular state.[171] Despite the volume of production of *Spirulina* worldwide, the same authors
452 reported that the study on vitamin A bioavailability in humans is limited and carried out using
453 *Spirulina* supplements.

454 The limitation of bioavailability of astaxanthin from *Haematococcus* is due to the physical
455 barrier of cysts rich in astaxanthin. Mechanical disruption and autoclaving of the astaxanthin-rich
456 algal biomass were effective to increase the pigment bioavailability.[172] The bioavailability of
457 astaxanthin from *Haematococcus* was related to the timing of ingestion; so, the astaxanthin
458 bioavailability was higher after meal than before meal.[173]

459 In summary, to produce a microalgal-based product that effectively has beneficial properties,
460 various aspects along the production process have to be taken into account. First, the cultivation
461 with optimal nutrients concentration or in starving condition affects the type of fatty acids present in
462 lipids and it appears to modulate the PUFAs position in the triacylglycerols with an impact on
463 bioavailability. On the other hand, the starvation reduces the proteins content; therefore, the
464 cultivation techniques should guarantee a balance between protein and lipid production. Second, it
465 is important to know the structure of the chemical compounds present in microalgae since this
466 affects the release of the different compounds making them available. Connected to this aspect is
467 the need to treat the biomass by different methods to aid the bioavailability of the valuable
468 intracellular compounds. Also important is the food matrix incorporating the microalgae since often
469 the presence of lipids in the food increases the bioavailability of valuable compounds such as
470 vitamins.

471

472 **Challenges and future perspectives**

473 The microalgal-based food and supplement/nutraceuticals global market is a sector with a great
474 potential of expansion. Currently, microalgae are used both as dried whole algae and to extract
475 high-value compounds to be used as ingredients, supplements, and colorants. Compared to the
476 market based on other sources, the microalgae volume of production and market size for food and
477 nutraceuticals are small with important sign of increase since the beginning of the century.[51] The
478 large-scale commercial production of microalgae for protein/carbohydrates impacting on food-feed
479 security can still be considered a less developed industry. Dried whole algae *Arthrospira* and
480 *Chlorella* for dietary supplements have the largest volume of production volumes worldwide (5000
481 and 2000 t of dry matter/year, respectively) with an estimated value of production of about \$40
482 millions/year each.[3] In addition, the market of high-value molecules from microalgae such as
483 astaxanthin, ω -3 fatty acids, and β -carotene has larger market potential in spite of smaller volume of
484 production. As example, the production from microalgae of polyunsaturated fatty acids such as
485 DHA and EPA is only 240 t/year with market value estimated to be higher than \$300
486 million/year.[3] The total production cost for microalgae-derived products becomes higher due to
487 the extraction techniques; for example, the total cost for microalgae astaxanthin is €465.58/kg.[3]
488 These values highlight that microalgal biomass is an expensive material compared to other types of
489 biomass (wheat straw 0.03 €/g).[174] For larger size plants (100–200 ha), the total cost (capital,
490 labor, other variables such as utilities and fertilizers) of microalgal biomass production, expressed
491 as kg dry weight, is estimated as 0.68 and 1.8€ for open ponds and flat photobioreactors (PBRs),
492 respectively. Considering the lower size of the real production plants, the resulting cost is higher
493 than the above estimation.[175,176] Recently, Ruiz et al.[177] reported differences in costs
494 according to the plants location; for a plant of 100 ha in South Spain, the total cost (capital and
495 operational costs) of microalgal biomass production is estimated 5.2 and 3.4 €/kg for open ponds

496 and flat PBRs, respectively, while for The Netherlands, these values are 11.0 and 6.0 €/kg. The
497 biorefinery cost (capital and operational costs) for food-feed sector is 0.9 and 1.4 €/kg for South
498 Spain and The Netherlands, respectively.

499 The current volume of market for *D. salina* is about 3000 t/year (for carotene), for *H. pluvialis*
500 is about 700 t (for astaxanthin), for *Shizochytrium* and *C. cohnii* is 20 and 500 t/year, respectively
501 (for DHA).[178] The volume of the microalgae market is expected to increase reaching US\$44.6
502 billion by 2023 with expanding at a compound annual growth rate of more than 5.2% from 2016 to
503 2023. Also, prevision of expansion is reported for the global nutraceutical market[178]; so, it is
504 foreseeable an increase in demand for microalgal biomass and valuable compounds.

505 In order to expand the industry of food products or ingredients based on microalgae, different
506 challenges need to be overcome. First, it is important to use a cultivation system highly efficient and
507 able to minimize contamination risks; this is a closed system in comparison to an open system
508 subjected to the weather variability and contamination risks. However, in large-scale cultivation
509 with closed systems, temperature and oxygen control are important to avoid culture collapsing.
510 Temperature control can be realized by water spraying on the PBRs or by heat exchangers while
511 oxygen control by using efficient degassers.[177] An efficient use of the light can be obtained by
512 correct design of the PBRs, the use of light-emitting diodes, or embedding light guides to
513 PBRs.[179,180] Moreover, the material of the PBRs has to prevent the microalgae adhesion to
514 avoid biofouling problems.[181] Another aspect to consider for an advantageous cultivation is the
515 integration in the microalgae plants of the systems to reuse CO₂ waste and waste streams rich in
516 phosphorous and nitrogen. In order to have a cost-effective scale-up, researches on PBRs are
517 needed. A way to improve the production of microalgae and reduce costs is growing the microalgae
518 in heterotrophic or mixotrophic cultivation. In particular, in mixotrophic process, microalgae
519 synthesize compounds through both autotrophic and heterotrophic pathways requiring less light
520 energy and organic compounds and, at the same time, recycling wastewater.[182] Another factor to

521 consider in order to reduce the cost of production is the location of the plants including the
522 microalgae ocean cultivation.[182] With the aim to monitor with the smallest possible delay the
523 process variables during the microalgae cultivation, new monitoring methods such as ion-specific
524 field effect transistor sensor for physicochemical parameters, biomass concentration by image
525 analysis, lipid concentration by infrared spectrophotometry, and dielectric scattering are
526 proposed.[183]

527 The biorefinery step involves costs for disrupting the microalgal biomass and for extracting
528 valuable compounds as well. Alternative methods of the current methods to disrupt cells such as
529 hydrodynamic cavitation, electrical fields, electromagnetic fields, and acoustic cavitation were
530 reported.[177] Usually, microalgae are produced for one specific aim, for example, to extract a
531 valuable pigment. A potentially more efficient approach could be a cascade biorefinery system that
532 allows the use of all the possible microalgal compounds.[167]

533 Another aspect is the selection of robust microalgal strains with increased tolerance to
534 photoinhibition, photosaturation, oxygen saturation, capture and conversion of CO₂, and producing
535 high concentration of compounds useful for the food/nutraceutical sector. Omics technologies are
536 also required to study pathways involved in the valuable compounds production and to understand
537 the role of nutrient depletion and other stress on their production.[184] On the other hand, a
538 possibility is the development and use of genetic-modified microalgae, but this requires
539 consideration of both the regulations about the use of modified organisms and the consumers'
540 acceptance.

541 Defining the best food formulation and the best way of food ingestion (raw or cooked food)
542 appears of great importance to enhance the bioavailability of the microalgal functional compounds.
543 This presupposes collaboration among phycologists, food engineers, and clinicians to rigorously
544 verify the compounds bioavailability.

545

546 The consumer acceptance of formulated new food plays an important role on their success on
547 the market. The acceptance or rejection of novel food depends on different factors[185] such as
548 sociodemographic factors, safety and healthiness perception of novel food, sensory appeal (taste,
549 texture, color, and odor), and consumer's lifestyle. To increase the microalgae-based
550 food/beverages market, it is necessary that consumers trust the novel formulates; therefore, a key
551 factor is the increase of consumers' knowledge about the microalgae used, the possible risks related
552 to their use, the nutritional and health benefits, and the technology of production.

553

554 **Conclusion**

555 Considering the consumer's demand for novel foods both healthier and sensorial attractive, the
556 modern food industry is oriented to design and produce new food formulations using natural
557 compounds. In this scenario, microalgae have acquired great interest since they confer to the
558 product improved nutritional values, functional properties other than attractiveness.

559 In the recent years, many investigations on the possible use of different microalgae –biomass
560 and metabolites – in a variety of foods have been carried out. In addition, various microalgae-based
561 products are available on the market all over the world even if the volume of production of these
562 foods remains limited.

563 To expand the sector of the microalgal fortified/based food and beverages, future needs
564 concern: microbiologists for discovering new microalgae species and their properties, engineers for
565 developing more cost-effective cultivation and biorefinery systems, food scientists for improving
566 sensory appeal, and clinicians for improving compounds availability.

567 The literature available on this field and the food and beverages currently available on the
568 market give the idea of the great interest for this sector and stimulate the different areas of expertise

569 in studying the microalgae, reducing cost of production, and designing new functional foods and
570 beverages.

571

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575

576 **References**

577 [1] Bordignon, G.; Cabrini, M. Algae: An Introduction. In *Algae as a Potential Source of Food and*
578 *Energy in Developing Countries Sustainability, Technology and Selected Case Studies*; Perosa, A.,
579 Bordignon, G., Ravagnan, G., Zinoviev, S., Eds.; Digital Publishing: Ca' Foscari Venezia, 2015; pp
580 9–17.

581 [2] Mata, T. M.; Martins, A. A.; Caetano, N. S. Microalgae for Biodiesel Production and Other
582 Applications: A Review. *Renew. Sustain. Ener. Rev.* 2010, 14, 217–232. DOI:
583 10.1016/j.rser.2009.07.020.

584 [3] Enzing, C.; Ploeg, M.; Barbosa, M.; Sijtsma, L. Microalgae-Based Products for the Food and
585 Feed Sector: An Outlook for Europe. In *JRC Scientific and Policy Reports, Report EUR 26255 EN*;
586 Vigani, M., Parisi, C., Rodríguez Cerezo, E., Eds.; Publications Office of the European Union:
587 Luxembourg, 2014; pp 1–82.

588 [4] Olaizola, M. Commercial Development of Microalgal Biotechnology: From the Test Tube to the
589 Marketplace. *Biomol. Eng.* 2003, 20, 459–466. DOI: 10.1016/S1389-0344(03)00076-5.

- 590 [5] Chacón -Lee, T. L.; González-Mariño, G. E. Microalgae for Healthy Foods: Possibilities and
591 Challenges. *Comp. Rev. Food Sci. Food. Safety*. 2010, 9, 655–675. DOI: 10.1111/j.1541-
592 4337.2010.00132.x.
- 593 [6] Borowitzka, M. A. High-Value Products from Microalgae - Their Development and
594 Commercialization. *J. Appl. Phycol.* 2013, 25, 743–756. DOI: 10.1007/s10811-013-9983-9.
- 595 [7] Dragone, G.; Fernandes, B.; Vicente, A. A.; Teixeira, J. A. Third Generation Biofuels from
596 Microalgae. In *Current Research Technology and Education Topics in Applied Microbiology and*
597 *Microbial Biotechnology*; Méndez-Vilas, A., Ed.; Formatex: Badajoz, Spain, 2010; pp 1355–1366.
- 598 [8] Duong, V. T.; Li, Y.; Nowak, E.; Schenk, P. M. Microalgae Isolation and Selection for
599 Prospective Biodiesel Production. *Energies*. 2012, 5, 1835–1849. DOI: 10.3390/en5061835.
- 600 [9] Lim, D. K. Y.; Garg, S.; Timmins, M.; Zhang, E. S. B.; Thomas-Hall, S. R.; Schuhmann, H.; Li,
601 Y.; Schenk, P. M. Isolation and Evaluation of Oil-Producing Microalgae from Subtropical Coastal
602 and Brackish Waters. *PloS. ONE*. 2012, 7, e40751. DOI: 10.1371/journal.pone.0040751.
- 603 [10] Pandey, R. K.; Kundu, K.; Prakash, V.; Bhaskar, H.; Karmakar, R.; Dahake, V. R. Production
604 of Biodiesel from Mixed Algal Culture and Its Fuel Characterization. *Int. J. Recent Sci. Res.* 2013,
605 4, 794–797.
- 606 [11] Sidari, R.; A Simple and Rapid Method for Separation and Isolation of Marine Algal Species
607 from Naturally Evolved Populations. *Mar. Biol. Res.* 2016, 12, 193–199. DOI:
608 10.1080/17451000.2015.1125003.
- 609 [12] Sidari, R.; Zema, D. A.; Bombino, G.; Sgrò, A.; Caridi, A. Evaluation of Natural Mixed Micro-
610 Algal Blooms as Potential Feedstock for Biodiesel Production. *Proceeding 23rdEUBCE 2015*. 231–
611 234, ISBN: 978-88-89407-516, Wien (Austria), 1-4 June 2015, doi:10.5071/23rdEUBCE2015-
612 1CV.1.61 2015.

- 613 [13] de Morais, M. G.; Da SilvaVaz, B.; de Morais, E. G.; Vieira Costa, J. A., Biologically Active
614 Metabolites Synthesized by Microalgae, *BioMed. Res. Int*, 2015, 2015, Article ID 835761, 15
615 pages.
- 616 [14] Pulz, O.; Gross, W. Valuable Products from Biotechnology of Microalgae. *Appl. Microbiol.*
617 *Biotechnol.* 2004, 65, 635–648. DOI: 10.1007/s00253-004-1647-x.
- 618 [15] Draaisma, R. B.; Wijffels, R. H.; Slegers, P. M.; Brentner, L. B.; Roy, A.; Barbosa, M. J. Food
619 Commodities from Microalgae. *Curr. Opin. Biotechnol.* 2013, 24, 169–177. DOI:
620 10.1016/j.copbio.2012.09.012.
- 621 [16] Hoseini, S. M.; Shahbazizadeh, S.; Khosravi-Darani, K.; Mozafari, M. R. *Spirulina Platensis*:
622 Food and Function. *Current. Nutr. Food Sci.* 2013a, 9, 189–193. DOI:
623 10.2174/1573401311309030003.
- 624 [17] Gouveia, L.; Batista, A. P.; Sousa, I.; Raymundo, A.; Bandarra, N. M. Microalgae in Novel
625 Food Products. In *Food Chemistry Research Development*; Papadopoulos, K.N., Ed.; Nova Science
626 Publishers, Inc.: New York, USA, Chapter 2, 2008a; pp 1–36.
- 627 [18] Laza-Martínez, A.; Fernández-Marín, B.; García-Plazaola, J. I. Rapid Colour Changes in
628 *Euglena Sanguinea* (Euglenophyceae) Caused by Internal Lipid Globule Migration. *Eur. J. Phycol.*
629 2019, 54, 91–101. DOI: 10.1080/09670262.2018.1513571.
- 630 [19] Priyadarshani, I.; Rath, B. Commercial and Industrial Applications of Microalgae – A Review.
631 *J. Algal. Biomass. Utln.* 2012, 3, 89–100.
- 632 [20] Spolaore, P.; Joannis-Cassan, C.; Duran, E.; Isambert, A. Commercial Applications of
633 Microalgae. *J. Biosci. Bioeng.* 2006, 101, 87–96. DOI: 10.1263/jbb.101.166.

- 634 [21] Costa, J. A. C.; de Moraes, M. G. Microalgae for Food Production. In Fermentation Process
635 Engineering in the Food Industry; Soccol, C.R., Pandey, A., Larroche, C., Eds.; Taylor & Francis:
636 UK, 2013; p 486.
- 637 [22] Barrow, C.; Shahidi, F. Marine Nutraceuticals and Functional Foods; Taylor & Francis Group:
638 UK, 2008.
- 639 [23] Ibañez, E.; Cifuentes, A. Benefits of Using Algae as Natural Sources of Functional Ingredients.
640 J. Sci. Food. Agr. 2013, 93, 703–709. DOI: 10.1002/jsfa.6023.
- 641 [24] Hoseini, S. M.; Khosravi-Darani, K.; Mozafari, M. R. Nutritional and Medical Applications of
642 *Spirulina* Microalgae. Mini. Rev. Med. Chem. 2013b, 13, 1231–1237. DOI:
643 10.2174/1389557511313080009.
- 644 [25] Sidhu, K. S.; Health Benefits and Potential Risks Related to Consumption of Fish or Fish Oil.
645 Reg. Toxicol. Pharmacol. 2003, 38, 336–344. DOI: 10.1016/j.yrtph.2003.07.002.
- 646 [26] Thies, F.; Garry, J. M.; Yagoob, P.; Rerkasem, K.; Williams, J.; Shearman, C. P.; Gallagher, P.
647 J.; Calder, P. C.; Grimble, R. F. Association of N-3 Polyunsaturated Fatty Acids with Stability of
648 Atherosclerotic Plaques: A Randomized Controlled Trial. Lancet. 2003, 361, 477–485. DOI:
649 10.1016/S0140-6736(03)12468-3.
- 650 [27] Seto, A.; Wang, H. L.; Hesseltine, C. W. Culture Conditions Affect Eicosapentaenoic Acid
651 Content of *Chlorella Minutissima*. J. Am. Oil Chem. Soc. 1984, 61, 892–894. DOI:
652 10.1007/BF02542159.
- 653 [28] Mahajan, G.; Kamat, M. γ -Linolenic Production from *Spirulina Platensis*. Appl. Microbiol.
654 Biotechnol. 1995, 43, 466–469. DOI: 10.1007/BF00218450.

- 655 [29] Chini Zittelli, G.; Lavista, F.; Bastianini, A.; Rodolfi, L.; Vincenzini, M.; Tredici, M. R.
656 Production of Eicosapentaenoic Acid by *Nannochloropsis* Sp. Cultures in Outdoor Tubular
657 Photobioreactors. *J. Biotechnol.* 1999, 70, 299–312. DOI: 10.1016/S0168-1656(99)00082-6.
- 658 [30] Bandarra, N. M.; Pereira, P. A.; Batista, I.; Vilela, M. H. Fatty acids, sterols and α -tocopherol
659 in *Isochrysis galbana*. *J. Food Lipids.* 2003, 18, 25–34. DOI: 10.1111/j.1745-4522.2003.tb00003.x.
- 660 [31] Donato, M.; Vilela, M. H.; Bandarra, N. M. Fatty Acids, Sterols, α -tocopherol and Total
661 Carotenoids Composition of *Diacronema Vlkianum*. *J. Food Lipids.* 2003, 10, 267–276.
662 DOI:10.1111/j.1745-4522.2003.tb00020.x.
- 663 [32] Mendes, A.; Reis, A.; Vasconcelos, R.; Guerra, P.; Lopes Da Silva, T. *Cryptocodinium*
664 *Cohnii* with Emphasis on DHA Production: A Review. *J. Appl. Phycol.* 2009, 21, 199–214.
665 DOI:10.1007/s10811-008-9351-3.
- 666 [33] Winwood, R. J.;. Recent Developments in the Commercial Production of DHA and EPA Rich
667 Oils from Micro-Algae. *Ocl.* 2013, 20(D604), 1–5. DOI: 10.1051/ocl/2013030.
- 668 [34] Ratledge, C.; Anderson, A.J.; Kanagachandran, K.; Grantham, D.; Stephenson, J.C.; de Swaaf,
669 M.; Sijtsma, L. Culture of *Cryptocodinium Cohnii* for the Synthesis of a Polyunsaturated Fatty
670 Acid. WO Patent 2001004338, 2001.
- 671 [35] Kulczyński, B.; Gramza-Michałowska, A.; Kobus-Cisowska, J.; Kmiecik, D. The Role of
672 Carotenoids in the Prevention and Treatment of Cardiovascular Disease – Current State of
673 Knowledge. *J. Func. Foods.* 2017, 38, 45–65. DOI: 10.1016/j.jff.2017.09.001.
- 674 [36] Lorenz, R. T.; Cysewski, G. R. Commercial Potential for *Haematococcus* Microalgae as a
675 Natural Source of Astaxanthin. *Trends. Biotechnol.* 2000, 18, 160–167. DOI: 10.1016/S0167-
676 7799(00)01433-5.

677 [37] Guerin, M.; Huntley, M. E.; Olaizola, M. *Haematococcus* Astaxanthin: Applications for
678 Human Health and Nutrition. *Trends. Biotechnol.* 2003, 21, 210–216. DOI: 10.1016/S0167-
679 7799(03)00078-7.

680 [38] Dufossé, L.; Galaup, P.; Yarnon, A.; Arad, S. M.; Blanc, P.; Chidambara Murthy, K. N.;
681 Ravishankar, G. A. Microorganisms and Microalgae as Source of Pigments for Use: A Scientific
682 Oddity or an Industrial Reality? *Trends Food Sci. Tech.* 2005, 16, 389–406.
683 DOI: 10.1016/j.tifs.2005.02.006.

684 [39] Del Campo, J. A.; Rodriguez, H.; Moreno, J.; Vargas, M. A.; Rivas, J.; Guerrero, M. G. Lutein
685 Production by *Muriellopsis* Sp. In an Outdoor Tubular Photobioreactor. *J. Biotechnol.* 2001, 85,
686 289–295. DOI: 10.1016/S0168-1656(00)00380-1.

687 [40] Del Campo, J. A.; García-González, M.; Guerrero, M. G. Outdoor Cultivation of Microalgae
688 for Carotenoid Production: Current State and Perspectives. *Appl. Microbiol. Biotechnol.* 2007, 74,
689 1163–1174. DOI: 10.1007/s00253-007-0844-9.

690 [41] García-González, M.; Moreno, J.; Manzano, J. C.; Florencio, F. J.; Guerrero, M. G. Production
691 of *Dunaliella Salina* Biomass Rich in 9-cis- β -carotene and Lutein in a Closed Tubular
692 Photobioreactor. *J. Biotechnol.* 2005, 115, 81–90. DOI: 10.1016/j.jbiotec.2004.07.010.

693 [42] Pangestuti, R.; Kim, S.-K. Biological Activities and Health Benefit Effects of Natural pigments
694 Derived from Marine Algae. *J. Funct. Foods.* 2011, 3, 255–266. DOI: 10.1016/j.jff.2011.07.001.

695 [43] Viskari, P. J.; Colyer, C. L. Rapid Extraction of Phycobiliproteins from Cultured
696 Cyanobacteria Samples. *Anal. Biochem.* 2003, 319, 263–271. DOI: 10.1016/S0003-
697 2697(03)00294-X.

698 [44] Sekar, S.; Chandramohan, M. Phycobiliproteins as a Commodity: Trends in Applied Research,
699 Patents and Commercialization. *J. Appl. Phycol.* 2008, 20, 113–136. DOI:10.1007/s10811-007-
700 9188-1.

701 [45] Gupta, A.; Sainis, J. K. Isolation of C-Phycocyanin from *Synechococcus* Sp., (*Anacystis*
702 *Nidulans* BD1). *J. Appl. Phycol.* 2010, 22, 231–233. DOI: 10.1007/s10811-009-9449-2.

703 [46] Román, R. B.; Álvarez-Pez, J. M.; Acién Fernández, F. G.; Molina Grima, E. Recovery of Pure
704 B-Phycoerythrin from the Microalga *Porphyridium Cruentum*. *J. Biotechnol.* 2002, 93, 73–85.

705 [47] Dupre, C.; Guary, J. C.; Grizeau, D. Culture of an Autoflocculent Microalga in a Vertical
706 Tubular Photobioreactor for Phycoerythrin Production. *Biotechnol. Tech.* 1995, 9, 185–190. DOI:
707 10.1007/BF00157076.

708 [48] Eriksen, N.;. Production of Phycocyanin - a Pigment with Applications in Biology,
709 Biotechnology, Foods and Medicine. *Appl. Microbiol. Biotechnol.* 2008, 80, 1–14.
710 DOI:10.1007/s00253-008-1535-x.

711 [49] Becker, W.;. Microalgae in Human and Animal Nutrition. In *Handbook of Microalgal Culture*;
712 Richmond, A., Ed.; Scientific Research – An Academic Publisher: Wuhan, 2004; pp 312–351.

713 [50] Vigani, M.; Parisi, C.; Rodríguez-Cerezo, E.; Barbosa, M. J.; Sijtsma, L.; Ploeg, M.; Enzing,
714 C. Food and Feed Products from Microalgae: Market Opportunities and Challenges for the EU.
715 *Trends Food Sci. Technol.* 2015, 42, 81–92. DOI: 10.1016/j.tifs.2014.12.004.

716 [51] Ismail, A.;. Marine Lipids Overview: Markets, Regulation, and the Value Chain. *Ocl.* 2010, 17,
717 205–208. DOI: 10.1051/ocl.2010.0321.

718 [52] Wells, M. L.; Potin, P.; Craigie, J. S.; Raven, J. A.; Merchant, S. S.; Helliwell, K. E.; Smith, A.
719 G.; Camire, M. E.; Brawley, S. H. Algae as Nutritional and Functional Food Sources: Revisiting
720 Our Understanding. *J. Appl. Phycol.* 2017, 29, 949–982.

721 [53] Brown, M. R.; The Amino Acid and Sugar Composition of 16 Species of Microalgae Used in
722 Mariculture. *J. Exp. Mar. Biol. Ecol.* 1991, 145, 79–99. DOI: 10.1016/0022-0981(91)90007-J.

723 [54] Becker, E. W.; *Microalgae: Biotechnology and Microbiology*; Cambridge University
724 Press:Cambridge, 1994.

725 [55] Zhu, C. J.; Lee, Y. K. Determination of Biomass Dry Weight of Marine Microalgae. *J. Appl.*
726 *Phycol.* 1997, 9, 189–194. DOI: 10.1023/A:1007914806640.

727 [56] Pleissner, D.; Eriksen, N. T. Effects of Phosphorous, Nitrogen, and Carbon Limitation on
728 Biomass Composition in Batch and Continuous Flow Cultures of the Heterotrophic Dinoflagellate
729 *Cryptocodinium Cohnii*. *Biotech. Bioeng.* 2012, 109(8), 2006–2016. DOI:10.1002/bit.24470.

730 [57] Ludevese-Pascual, G.; Dela Peña, M.; Tornalejo, J. Biomass Production, Proximate
731 Composition and Fatty Acid Profile of the Local Marine Thraustochytrid Isolate, *Schizochytrium*
732 *Sp. LEY7* Using Low-Cost Substrates at Optimum Culture Conditions. *Aquacult. Res.* 2016, 47,
733 318–328. DOI: 10.1111/are.12494.

734 [58] Barka, A.; Blecker, C. Microalgae as A Potential Source of Single-Cell Proteins. A Review.
735 *Biotechnol. Agron. Soc. Environ.* 2016, 20, 427–436.

736 [59] Kent, M.; Welladsen, H. M.; Mangott, A.; Li, Y. Nutritional Evaluation of Australian
737 Microalgae as Potential Human Health Supplements. *PloS. ONE.* 2015, 10, e0118985.
738 DOI:10.1371/journal.pone.0118985.

739 [60] Brown, M. R.; Garland, C. D.; Jeffrey, S. W.; Jameson, I. D.; Leroi, J. M. The Gross and
740 Amino Acid Compositions of Batch and Semi-Continuous Cultures of *Isochrysis Sp.* (Clone
741 T.ISO), *Pavlova Lutheri* and *Nannochloropsis Oculata*. *J. Appl. Phycol.* 1993, 5, 285–296.
742 DOI:10.1007/BF02186231.

- 743 [61] Brown, M. R.; Jeffrey, S. W. The Amino Acid and Gross Composition of Marine Diatoms
744 Potentially Useful for Mariculture. *J. Appl. Phycol.* 1995, 7, 521–527. DOI: 10.1007/BF00003938.
- 745 [62] Skrede, A.; Mydland, L. T.; Ahlstrøm, Ø.; Reitan, K. I.; Gislerød, H. R.; Øverland, M.
746 Evaluation of Microalgae as Sources of Digestible Nutrients for Monogastric Animals. *J. Animal.*
747 *Feed. Sci.* 2011, 20, 131–142. DOI: 10.22358/jafs/66164/2011.
- 748 [63] Tibbetts, S. M.; Milley, J. E.; Lall, S. P. Chemical Composition and Nutritional Properties of
749 Freshwater and Marine Microalgal Biomass Cultured in Photobioreactors. *J. Appl. Phycol.* 2015,
750 27, 1109–1119. DOI: 10.1007/s10811-014-0428-x.
- 751 [64] Madeira, M. S.; Cardoso, C.; Lopes, P. A.; Coelho, D.; Afonso, C.; Bandarra, N. M.; Pratesa, J.
752 A. M. Microalgae as Feed Ingredients for Livestock Production and Meat Quality: A Review.
753 *Livest. Sci.* 2017, 205, 111–121. DOI: 10.1016/j.livsci.2017.09.020.
- 754 [65] Patil, V.; Källqvist, T.; Olsen, E.; Vogt, G.; Gislerød, H. R. Fatty Acid Composition of 12
755 Microalgae for Possible Use in Aquaculture Feed. *Aquacult. Int.* 2007, 15, 1–9. DOI:
756 10.1007/s10499-006-9060-3.
- 757 [66] Martins, D. A.; Custódio, L.; Barreira, L.; Pereira, H.; Ben-Hamadou, R.; Varela, J.; Abu-
758 Salah, K. M. Alternative Sources of N-3 Long-Chain Polyunsaturated Fatty Acids in Marine
759 Microalgae. *Mar. Drugs.* 2013, 11, 2259–2281. DOI: 10.3390/md11072259.
- 760 [67] Renaud, S. M.; Parry, D. L.; Thinh, L.-V. Microalgae for Use in Tropical Aquaculture I: Gross
761 Chemical and Fatty Acid Composition of Twelve Species of Microalgae from the Northern
762 Territory, Australia. *J. Appl. Phycol.* 1994, 6, 337–345. DOI: 10.1007/BF02181948.
- 763 [68] Hu, H.; Yeguang, L.; Chuntao, Y.; Yexin, O. Isolation and Characterization of a Mesophilic
764 *Arthrospira Maxima* Strain Capable of Producing Docosahexaenoic Acid. *J. Microbiol. Biotechnol.*
765 2011, 21(7), 697–702. DOI: 10.4014/jmb.1101.12040.

766 [69] Ötles, S.; Pire, R. Fatty Acid Composition of *Chlorella* and *Spirulina* Microalgae Species. J.
767 AOAC. Int. 2001, 84(6), 1708–1714.

768 [70] Markou, G.; Nerantzis, E. Microalgae for High-Value Compounds and Biofuels Production: A
769 Review with Focus on Cultivation under Stress Conditions. Biotechnol. Adv. 2013, 31, 1532–1542.
770 DOI: 10.1016/j.biotechadv.2013.07.011.

771 [71] Orosa, M.; Franqueira, D.; Cid, A.; Abalde, J. Carotenoid Accumulation in *Haematococcus*
772 *Pluvialis* in Mixotrophic Growth. Biotech. Lett. 2001, 23(5), 373–378. DOI:
773 10.1023/A:1005624005229.

774 [72] Abd El-Baky, H. H.; El Baz, F. K.; El-Baroty, G. S. *Spirulina* Species as a Source of
775 Carotenoids and α -tocopherol and Its Anticarcinoma Factors. Biotechnol. 2003, 2(3), 222–240.
776 DOI: 10.3923/biotech.2003.222.240.

777 [73] Abd El-Baky, H. H.; El Baz, F. K.; El-Baroty, G. S. Production of Antioxidant by the Green
778 Alga *Dunaliella Salina*. Int. J. Agri. Biol. 2004, 6(1), 49–57.

779 [74] Del Campo, J. A.; Rodríguez, H.; Moreno, J.; Vargas, M. A.; Rivas, J.; Guerrero, M. G.
780 Accumulation of Astaxanthin and Lutein in *Chlorella Zofingiensis* (Chlorophyta). Appl. Microbiol.
781 Biotechnol. 2004, 64(6), 848–854. DOI: 10.1007/s00253-003-1490-5.

782 [75] Di Sanzo, G.; Mehariya, S.; Martino, M.; Larocca, V.; Casella, P.; Chianese, S.; Musmarra, D.;
783 Balducci, R.; Molino, A. Supercritical Carbon Dioxide Extraction of Astaxanthin, Lutein, and
784 Fatty Acids from *Haematococcus Pluvialis* Microalgae. Mar. Drugs. 2018, 16(334), 2–18. DOI:
785 10.3390/md16090334.

786 [76] Sathasivam, R.; Ki, J.-S. A Review of the Biological Activities of Microalgal Carotenoids and
787 Their Potential Use in Healthcare and Cosmetic Industries. Mar. Drugs. 2018, 16(26), 2–31. DOI:
788 10.3390/md16010026.

789 [77] Fabregas, J.; Herrero, C. Vitamin Content of Four Marine Microalgae. Potential Use as Source
790 of Vitamins in Nutrition. *J. Ind. Microbiol.* 1990, 5, 259–264. DOI: 10.1007/BF01569683.

791 [78] Hadley, K. B.; Bauer, J.; Milgram, N. W. The Oil-Rich Alga *Schizochytrium* Sp. As a Dietary
792 Source of Docosahexaenoic Acid Improves Shape Discrimination Learning Associated with Visual
793 Processing in a Canine Model of Senescence. *Prostag. Leukotr. ESS.* 2017, 118, 10–18. DOI:
794 10.1016/j.plefa.2017.01.011.

795 [79] Andrade, L. M.; Andrade, C. J.; Dias, M.; Nascimento, C. A. O.; Mendes, M. A. *Chlorella* and
796 *Spirulina* Microalgae as Sources of Functional Foods, Nutraceuticals, and Food Supplements; an
797 Overview. *MOJ. Food. Process. Technol.* 2018, 6(1), 00144. DOI: 10.15406/mojfpt.2018.06.00144.

798 [80] Regulation (EC) N° 258/97 of the European Parliament and of the 27 January 1991 concerning
799 novel foods and novel food ingredients.

800 [81] Champenois, J.; Marfaing, H.; Pierre, R. Review of the Taxonomic Revision of *Chlorella* and
801 Consequences for Its Food Uses in Europe. *J. Appl. Phycol.* 2015, 27, 1845–1851.
802 DOI:10.1007/s10811-014-0431-2.

803 [82] Commission Implementing Decision (EU) 2015/545 of 31 March 2015 authorising the placing
804 on the market of oil from the micro-algae *Schizochytrium* sp. (ATCC PTA-9695) as a novel food
805 ingredient under Regulation (EC) N° 258/97 of the European Parliament and of the Council
806 (notified under document C(2015) 2082), Brussels, Belgium.

807 [83] Commission Implementing Decision (EU) 2015/546 of 31 March 2015 authorising an
808 extension of use of DHA and EPA-rich oil from the micro-algae *Schizochytrium* sp. as a novel food
809 ingredient under Regulation (EC) N° 258/97 of the European Parliament and of the Council
810 (notified under document C(2015) 2083), Brussels, Belgium.

811 [84] Commission Implementing Decision 2014/463/EU: on authorising the placing on the market of
812 oil from the micro-algae *Schizochytrium* sp. as a novel food ingredient under Regulation (EC) N°
813 258/97 of the European Parliament and of the Council and repealing Decisions 2003/427/EC and
814 2009/778/EC, Brussels, Belgium.

815 [85] Commission Decision 2009/777/EC of 21 October 2009 concerning the extension of uses of
816 algal oil from the micro-algae *Ulkenia* sp. as a novel food ingredient under Regulation (EC) N°
817 258/97 of the European Parliament and of the Council (notified under document C(2009) 7932),
818 Brussels, Belgium.

819 [86] Regulation (EU) 2015/2283 of the European Parliament and of the Council of 25 November
820 2015 on novel foods, amending Regulation (EU) N° 1169/2011 of the European Parliament and of
821 the Council and repealing Regulation (EC) N° 258/97 of the European Parliament and of the
822 Council and Commission Regulation (EC) N° 1852/2001, Brussels, Belgium.

823 [87] GRAS Notice N° 000101 (2002) - GRAS Notification for *Spirulina* microalgae, Maryland,
824 USA.

825 [88] JETRO Specifications and Standards for Foods, Food Additives, etc. Under the Food
826 Sanitation Act, Japan External Trade Organization (JETRO): Tokyo, Japan, 2011, 1–180.

827 [89] GRAS Notice N° 000137 (2003) - GRAS Exemption claim for DHA algal oil derived from
828 *Schizochytrium* sp. as a source of DHA for use in foods, Maryland, USA.

829 [90] GRAS Notice N° 000160 (2004) - Expert panel consensus oil under the conditions of intended
830 use in traditional foods, Maryland, USA.

831 [91] GRAS Notice N° 000330 (2010) - GRAS Notice for an Algal flour (*Chlorella*) ingredient,
832 Maryland, USA.

833 [92] GRAS Notice N° 000351 (2010) - GRAS Assessment for Nikken Sohonsa Corporation
834 *Dunaliella bardawil* powder, Maryland, USA.

- 835 [93] GRAS Notice N° 000396 (2011) - GRAS exemption claim for *Chlorella vulgaris* as an
836 ingredient in foods, Maryland, USA.
- 837 [94] GRAS Notice N° 697 (2017) - GRAS Notice for dried *Euglena gracilis* (ATCC PTA-123017),
838 Maryland, USA.
- 839 [95] Australia New Zealand Food Standards Code – Schedule 25 – Permitted novel foods.
840 Authorised Version F2017C00413 registered 25/05/2017, Canberra (Australia), Wellington (New
841 Zealand), pp 1–3.
- 842 [96] Guinee, T. P.; Caric, M.; Kaláb, M. Pasteurized Processed Cheese and Substitute/Imitation
843 Cheese Products. In Cheese: Chemistry Physics and Microbiology. Major Cheese Groups; Fox,
844 P.F., McSweeney, P.L.H., Cogan, T.M., Guinee, T.P., Eds.; Elsevier Ltd: London, 2004; pp 349–
845 394.
- 846 [97] Heo, J. Y.; Shin, H. J.; Oh, D. H.; Cho, S. K.; Yang, C. J.; Kong, I. K.; Lee, S. S.; Choi, K. S.;
847 Choi, S. H.; Kim, S. C.; et al. Quality Properties of Appenzeller Cheese Added with *Chlorella*.
848 Korean. J. Food Sci. Animal. Res. 2006, 26, 525–531.
- 849 [98] Habib, M. A. B.; Parvin, M.; Huntington, T. C.; Hasan, M. R. A Review on Culture,
850 Production and Use of *Spirulina* as Food for Humans and Feeds for Domestic Animals and Fish.
851 FAO Fisheries and Aquaculture Circular No. 1034, 2008.
- 852 [99] Jeon, J.-K.; Effect of *Chlorella* Addition on the Quality of Processed Cheese. J. Korean Soc.
853 Food Sci. Nutr. 2006, 35, 373–377. DOI: 10.3746/jkfn.2006.35.3.373.
- 854 [100] Mohamed, A. G.; Abo-El-Khair, E.; Shalaby, S. M. Quality of Novel Healthy Processed
855 Cheese Analogue Enhanced with Marine Microalgae *Chlorella Vulgaris* Biomass. World. Appl.
856 Sci. J. 2013, 23, 914–925.

- 857 [101] Shalaby, S. M.; Yasin, N. M. N. Quality Characteristics of Croissant Stuffed with Imitation
858 Processed Cheese Containing Microalgae *Chlorella Vulgaris* Biomass. World J. Dairy. Food. Sci.
859 2013, 8, 58–66.
- 860 [102] Shirota, M.; Nagamatsu, N.; Takechi, Y. Method for Cultivating Lactobacilli. U.S. Pat. No.
861 3,123,538, 1964.
- 862 [103] Gibson, G. R.; Roberfroid, M. B. Dietary Modulation of the Human Colonic Microbiota:
863 Introducing the Concept of Prebiotics. J. Nutr. 1995, 125, 1401–1412. DOI: 10.1093/jn/125.6.1401.
- 864 [104] Parada, J. L.; de Caire, G. Z.; de Mulé, M. C. Z.; de Cano, M. M. S. Lactic Acid Bacteria
865 Growth Promoters from *Spirulina Platensis*. Int. J. Food. Microbiol. 1998, 45, 225–228.
866 DOI:10.1016/S0168-1605(98)00151-2.
- 867 [105] Gyenis, B.; Szigeti, J.; Molnár, N.; Varga, L. Use of Dried Microalgal Biomasses to Stimulate
868 Acid Production and Growth of *Lactobacillus Plantarum* and *Enterococcus Faecium* in Milk. Acta.
869 Agraria. Kaposváriensis. 2005, 9, 53–59.
- 870 [106] Molnár, N.; Gyenis, B.; Varga, L. Influence of Powdered *Spirulina Platensis* Biomass on
871 Acid Production of Lactococci in Milk. Milchwissenschaft. 2005, 4, 380–382.
- 872 [107] Molnár, N.; Sipos-Kozma, Z.; Tóth, Á.; Ásványi, B. M.; Varga, L. Development of
873 Functional Dairy Food Enriched in *Spirulina (Arthrospira Platensis)*. Tejgazdaság. 2009, 69, 15–
874 22.
- 875 [108] Guldás, M.; Irkin, R. Influence of *Spirulina Platensis* Powder on the Microflora of Yoghurt
876 and Acidophilus Milk. Mljekarstvo. 2010, 60, 237–243.
- 877 [109] Beheshtipour, H.; Mortazavian, A. M.; Haratian, P.; Khosravi-Darani, K. Effects of *Chlorella*
878 *Vulgaris* and *Spirulina Platensis* Addition on the Viability of Probiotic Bacteria in Yoghurt and Its
879 Biochemical Properties. Eur. Food Res. Technol. 2012, 235, 719–728. DOI: 10.1007/s00217-012-
880 1798-4.

- 881 [110] Beheshtipour, H.; Mortazavian, A. M.; Mohammadi, R.; Sohrabvandi, S.; Khosravi-Darani,
882 K. Supplementation of *Spirulina Platensis* and *Chlorella Vulgaris* Algae into Probiotic Fermented
883 Milks. *Comp. Rev. Food Sci. Food. Safety*. 2013, 12, 144–154. DOI: 10.1111/1541-4337.12004.
- 884 [111] Varga, L.; Sule, J.; Szigeti, J. Stimulation of Probiotic Lactobacilli and Bifidobacteria in
885 Cultured Dairy Foods. Proceedings of the International Scientific Conference on Sustainable
886 Development and Ecological Footprint. 26-27 March 2012, Sopron, Hungary. DOI: 10.1094/PDIS-
887 11-11-0999-PDN
- 888 [112] Prakash, D. R.; Pooja, K. Preparation of Low-Fat and High-Protein Frozen Yoghurt Enriched
889 with Papaya Pulp and *Spirulina*. *Trends. Biosci.* 2011, 4, 182–184.
- 890 [113] Cho, E. J.; Nam, E. S.; Park, S. I. Keeping Quality and Sensory Properties of Drinkable
891 Yoghurt with Added *Chlorella* Extract. *Korean. J. Food Nutr.* 2004, 17(2), 128–132.
- 892 [114] Kavimandan, A. Incorporation of *Spirulina Platensis* into Probiotic Fermented Dairy
893 Products. *Int. J. Dairy Sci.* 2015, 10, 1–11. DOI: 10.3923/ijds.2015.1.11.
- 894 [115] Ak, B.; Avşaroğlu, E.; Işık, O.; Özyurt, G.; Kafkas, E.; Etyemez, M.; Uslu, L. Nutritional and
895 Physicochemical Characteristics of Bread Enriched with Microalgae *Spirulina Platensis*. *Int. J. Eng.*
896 *Res. Appl.* 2016, 6, 30–38.
- 897 [116] Jeong, C. H.; Cho, H. J.; Shim, K. H. Quality Characteristics of White Bread Added with
898 *Chlorella* Powder. *Korean. J. Food Pres.* 2006, 13, 465–471.
- 899 [117] Babuskin, S.; Krishnan, K. R.; Babu, P. A. S.; Sivarajan, M. S.; Sukumar, M. Functional
900 Foods Enriched with Marine Microalga *Nannochloropsis Oculata* as a Source of ω -3 Fatty Acids.
901 *Food Technol. Biotechnol.* 2014, 52, 292–299.

- 902 [118] Gouveia, L.; Batista, A. P.; Miranda, A.; Empis, J.; Raymundo, A. *Chlorella Vulgaris*
903 Biomass Used as Colouring Source in Traditional Butter Cookies. *Innov. Food Sci. Emerg.*
904 *Technol.* 2007, 8, 433–436. DOI: 10.1016/j.ifset.2007.03.026.
- 905 [119] Bang, B.-H.; Kim, K.-P.; Jeong, E.-J. Quality Characteristics of Cookies that Contain
906 Different Amounts of *Chlorella* Powder. *Korean J. Food Preserv.* 2013, 20, 798–804. DOI:
907 10.11002/kjfp.2013.20.6.798.
- 908 [120] Gouveia, L.; Coutinho, C.; Mendonça, E.; Batista, A. P.; Sousa, I.; Bandarra, N. M.;
909 Raymundo, A. Sweet Biscuits with *Isochrysis Galbana* Microalga Biomass as a Functional
910 Ingredient. *J. Sci. Food. Agri.* 2008c, 88, 891–896. DOI: 10.1002/jsfa.3166.
- 911 [121] Salehifar, M.; Shahbazizadeh, S.; Khosravi-Darani, K.; Behmadi, H.; Ferdowsi, R. Possibility
912 of Using Microalgae *Spirulina Platensis* Powder in Industrial Production of Iranian Traditional
913 Cookies. *Iranian J. Nutr. Sci. Food Technol.* 2013, 7, 63–72.
- 914 [122] Lucas, B. F.; de Morais, M. G.; Santosa, T. D.; Costa, J. A. V. *Spirulina* for Snack
915 Enrichment: Nutritional, Physical and Sensory Evaluations. *LWT – Food. Sci. Technol.* 2018, 90,
916 270–276. DOI: 10.1016/j.lwt.2017.12.032.
- 917 [123] Gouveia, L.; Batista, A. P.; Raymundo, A.; Bandarra, N. M. *Spirulina Maxima* and
918 *Diacronema Vlkianum* Microalgae in Vegetable Gelled Desserts. *Nutr. Food Sci.* 2008b, 38, 492–
919 501. DOI: 10.1108/00346650810907010.
- 920 [124] Kim, K. J.; Chung, H. C. Quality Characteristics of Yellow Layer Cake Containing Different
921 Amounts of *Chlorella* Powder. *Korean J. Food Cook. Sci.* 2010, 26, 860–865. DOI:
922 10.5851/kosfa.2010.30.5.860.
- 923 [125] Fradique, M.; Batista, A. P.; Nunes, M. C.; Gouveia, L.; Bandarra, N. M.; Raymundo, A.
924 *Chlorella Vulgaris* and *Spirulina Maxima* Biomass Incorporation in Pasta Products. *J. Sci. Food*
925 *Agr.* 2010, 90, 1656–1664. DOI: 10.1002/jsfa.3999.

926 [126] Acheson, J.; *Chlorella* in Packed Food, Beverage, and Pet Products. Global Analysis Report.
927 Her Majesty the Queen in Right of Canada, represented by the Minister of Agriculture and Agri-
928 Food. 1–10, 2016.

929 [127] Gouveia, L.; Raymundo, A.; Batista, A. P.; Miranda, A.; Sousa, I.; Empis, J. Colouring
930 Emulsions Using Microbial Biomass – Stability over Time. In *Pigments in Food – More than*
931 *Colours*; Dufossé, L., Ed.; Pigments Publishing, Université de Bretagne Occidentale: Quimper,
932 2004; pp 121–123.

933 [128] Raymundo, A.; Gouveia, L.; Batista, A. P.; Empis, J.; Sousa, I. Fat Mimetic Capacity of
934 *Chlorella Vulgaris* Biomass in Oil-In-Water Food Emulsions Stabilized by Pea Protein. *Food Res.*
935 *Int.* 2005, 38, 961–965. DOI: 10.1016/j.foodres.2005.02.016.

936 [129] Gouveia, L.; Batista, A. P.; Raymundo, A.; Sousa, I.; Empis, J. *Chlorella Vulgaris* and
937 *Haematococcus Pluvialis* Biomass as Colouring and Antioxidant in Food Emulsions. *Eur. Food*
938 *Res. Technol.* 2006, 222, 362–367. DOI: 10.1007/s00217-005-0105-z.

939 [130] Batista, A. P.; Nunes, M. C.; Fradinho, P.; Gouveia, L.; Sousa, I.; Raymundo, A.; Franco, J.
940 M. Novel Foods with Microalgae Ingredients - Effect of Gel Setting Conditions on the Linear
941 Viscoelasticity of *Spirulina* and *Haematococcus* Gels. *J. Food. Eng.* 2012, 110, 182–189.
942 DOI:10.1016/j.jfoodeng.2011.05.044.

943 [131] Kim, D. C.; Won, S. I.; In, M.-J. Preparation and Quality Characteristics of Mul-Kimchi
944 Added with *Chlorella*. *J. Appl. Biol. Chem.* 2014, 57, 23–28. DOI: 10.3839/jabc.2014.004.

945 [132] Park, M.-K.; Lee, J.-M.; Park, C.-H.; In, M.-J. Quality Characteristic of Sulgidduk Containing
946 *Chlorella* Powder. *J. Korean. Soc. Food Sci. Nutr.* 2002, 31, 225–229. DOI:
947 10.3746/jkfn.2002.31.2.225.

948 [133] Jorgensen, J.; Fermentation Process for Producing Alcoholic Beverages from Microalgae.
949 United States Patent 3,389,998, 1968. DOI: 10.1055/s-0028-1105114

950 [134] Lone, J.; Lene, D. S.; Karsten, O.; Skibsted, L. H. Heat and Light Stability of Three Natural
951 Blue Colourants for Use in Confectionery and Beverages. *Eur. Food Res. Technol.* 2005, 220, 261–
952 266. DOI: 10.1007/s00217-004-1062-7.

953 [135] Zhongliang, L.; Changlan, Z. Blue Liquid Spirulina Beverage and Preparing Process Thereof.
954 CN Patent 1127611, 1996.

955 [136] Xu, X.; 1994 Natural Blue Beverage Preparing Method Made of Spiral Algae. CN Patent
956 1096178. DOI: 10.3168/jds.S0022-0302(94)77044-2

957 [137] Keillar, D.; Drink Containing Fluorescent Agent. WO Patent 03099039, 2003.

958 [138] Goiris, K.; Muylaert, K.; De Cooman, L. Microalgae as a Novel Source of Antioxidants for
959 Nutritional Applications. In *Handbook of Marine Microalgae: Biotechnology Advances*; Chapter
960 17; Kim, S.-K., Ed.; Academic Press, Elsevier: USA, 2015; pp 269–280.

961 [139] Valencia, I.; Ansorena, D.; Astiasarán, I. Development of Dry Fermented Sausages Rich in
962 Docosaehaenoic Acid with Oil from the Microalgae *Schizochytrium* Sp.: Influence on Nutritional
963 Properties, Sensorial Quality and Oxidation Stability. *Food. Chem.* 2007, 104, 1087–1096.
964 DOI:10.1016/j.foodchem.2007.01.021.

965 [140] García-Íñiguez de Ciriano, M.; Larequi, E.; Rehecho, S.; Calvo, M. I.; Cavero, R. Y.;
966 Navarro-Blasco, I.; Astiasarán, I.; Ansorena, D. Selenium, Iodine, W-3 PUFA and Natural
967 Antioxidant from *Melissa Officinalis* L.: A Combination of Components from Healthier Dry
968 Fermented Sausages Formulation. *Meat. Sci.* 2010, 85, 274–279. DOI:
969 10.1016/j.meatsci.2010.01.012.

970 [141] Chee, C. P.; Gallaher, J. J.; Djordjevic, D.; Coupland, J. N. Chemical and Sensory Analysis of
971 Strawberry Flavoured Yogurt Supplemented with an Algae Oil Emulsion. *J. Dairy. Res.* 2005, 72,
972 311–316. DOI: 10.1017/S0022029905001068.

973 [142] Chee, C. P.; Djordjevic, D.; Faraji, H.; Decker, E. A.; Hollender, R.; McClements, D. J.;
974 Peterson, D. G.; Roberts, R. F.; Coupland, J. N. Sensory Properties of Vanilla and Strawberry
975 Flavored Ice Cream Supplemented with Omega-3 Fatty Acids. *Milchwissenschaft*. 2007, 62, 66–69.

976 [143] Gallaher, J. J.; Hollender, D. G.; Peterson, R. F.; Coupland, J. N. Effect of Composition and
977 Antioxidants on the Oxidative Stability of Fluid Milk Supplemented with Algae Oil Emulsion. *Int.*
978 *Dairy. J.* 2005, 15, 333–341. DOI: 10.1016/j.idairyj.2004.08.010.

979 [144] FSANZ, 2002 – 09/02 Application A428 DHA-rich dried marine microalgae (*Schizochytrium*
980 sp.) and DHA-rich oil derived from *Schizochytrium* sp. as novel food ingredients, Canberra
981 (Australia), Wellington (New Zealand), pp 1–70.

982 [145] FSANZ, 2005 – 2/05 Application A522 DHA-rich micro-algal oil from *Ulkenia* sp. as a novel
983 food. pp 1–180.

984 [146] Food and Drug Administration: 2003 Guidance for Industry. Bioavailability and
985 Bioequivalence Studies for Orally Administered Drug Products – General Considerations.
986 <http://www.fda.gov/downloads/Drugs/./Guidances/ucm070124.pdf>.

987 [147] Bleakley, S.; Hayes, M. Algal Proteins: Extraction, Application, and Challenges Concerning
988 Production. *Foods*. 2017, 6, 1–34. DOI: 10.3390/foods6080062.

989 [148] Minekus, M.; Alminger, M.; Alvito, P.; Ballance, S.; Bohn, T.; Bourlieu, C.; Carriere, F.;
990 Boutrou, R.; Corredig, M.; Dupont, D. A Standardised Static in Vitro Digestion Method Suitable
991 for Food - an International Consensus. *Food Funct.* 2014, 5, 1113–1124.
992 DOI:10.1039/C3FO60702J.

993 [149] Egger, L.; Menard, O.; Delgado-Andrade, C.; Alvito, P.; Assunção, R.; Balance, S.; Barberá,
994 R.; Brodkorb, A.; Cattenoz, T.; Clemente, A. et al. The Harmonized Infogest in Vitro Digestion
995 Method: From Knowledge to Action. *Food Res. Int.* 2016, 88, 217–225. DOI:
996 10.1016/j.foodres.2015.12.006.

- 997 [150] Becker, E. Micro-Algae as a Source of Protein. *Biotechnol. Adv.* 2007, 25, 207–210.
998 DOI:10.1016/j.biotechadv.2006.11.002.
- 999 [151] Hedenskog, G.; Enebo, L.; Vendlova, J.; Prokes, B. Investigation of Some Methods for
1000 Increasing the Digestibility of Microalgae. *Biotechnol. Bioeng.* 1969, 11, 37–51.
1001 DOI:10.1002/bit.260110104.
- 1002 [152] Devi, M. A.; Subbulakshmi, G.; Devi, K. M.; Venkataraman, L. V. Studies on the Proteins of
1003 Mass-Cultivated Blu-Green Alga (*Spirulina Platensis*). *J. Agric. Food Chem.* 1981, 29, 522–525.
1004 DOI: 10.1021/jf00105a022.
- 1005 [153] Morris, H. J.; Almarales, A.; Carrillo, O.; Bermudez, R. C. Utilisation of *Chlorella Vulgaris*
1006 Cell Biomass for the Production of Enzymatic Protein Hydrolysates. *Bioresour. Technol.* 2008, 99,
1007 7723–7729. DOI: 10.1016/j.biortech.2008.01.080.
- 1008 [154] Tibbetts, S. M.; Milley, J. E.; Lall, S. P. Chemical Composition and Nutritional Properties of
1009 Freshwater and Marine Microalgal Biomass Cultured in Photobioreactors. *J. Appl. Phycol.* 2014,
1010 27, 1109–1119. DOI: 10.1007/s10811-014-0428-x.
- 1011 [155] Cavonius, L. R.; Albers, E.; Undeland, I. In Vitro Bioaccessibility of Proteins and Lipids of
1012 pH-shift Processed *Nannochloropsis Oculata* Microalga. *Food. Funct.* 2016, 7, 2016–2024.
1013 DOI:10.1039/C5FO01144B.
- 1014 [156] Hori, K.; Ueno-Mohri, T.; Okita, T.; Ishibashi, G. Chemical Composition, in Vitro Protein
1015 Digestibility and in Vitro Available Iron of Blue Green Alga, *Nostoc Commune*. *Plant Foods Hum.*
1016 *Nutr.* 1990, 40, 223–229. DOI: 10.1007/BF01104146.
- 1017 [157] Joubert, Y.; Fleurence, J. Simultaneous Extraction of Proteins and DNA by an Enzymatic
1018 Treatment of the Cell Wall of *Palmaria Palmata* (Rhodophyta). *J. Appl. Phycol.* 2008, 20, 55–61.
1019 DOI: 10.1007/s10811-007-9180-9.

- 1020 [158] Wong, K.; Cheung, P. C. Nutritional Evaluation of Some Subtropical Red and Green
1021 Seaweeds Part II. In Vitro Protein Digestibility and Amino Acid Profiles of Protein Concentrates.
1022 Food. Chem. 2001, 72, 11–17. DOI: 10.1016/S0308-8146(00)00176-X.
- 1023 [159] Řezanka, T.; Lukavský, J.; Nedbalová, L.; Sigler, K. Production of Structured
1024 Triacylglycerols from Microalgae. Phytochem. 2014, 104, 95–104. DOI:
1025 10.1016/j.phytochem.2014.04.013.
- 1026 [160] Kagan, M. L.; West, A. L.; Zante, C.; Calder, P. C. Acute Appearance of Fatty Acids in
1027 Human Plasma – A Comparative Study between Polar-Lipid Rich Oil from the Microalgae
1028 *Nannochloropsis Oculata* and Krill Oil in Healthy Young Males. Lipids Health Dis. 2013, 12, 102.
1029 DOI: 10.1186/1476-511X-12-102.
- 1030 [161] Hulatt, C. J.; Wijffels, R. H.; Bolla, S.; Kiron, V. Production of Fatty Acids and Protein by
1031 *Nannochloropsis* in Flat-Plate Photobioreactors. PLoS. ONE. 2017, 12, 1–17. DOI:
1032 10.1371/journal.pone.0170440.
- 1033 [162] Schuchardt, J. P.; Schneider, I.; Meyer, H.; Neubronner, J.; von Schacky, C.; Hahn, A.
1034 Incorporation of EPA and DHA into Plasma Phospholipids in Response to Different Omega-3 Fatty
1035 Acid Formulations - a Comparative Bioavailability Study of Fish Oil Vs. Krill Oil. Lipids. Health.
1036 Dis. 2011, 10, 145. DOI: 10.1186/1476-511X-10-232.
- 1037 [163] de Mello-Sampayo, C.; Paterna, A.; Polizzi, A.; Duarte, D.; Batista, I.; Pinto, R.; Gonçalves,
1038 P.; Raymundo, A.; Batista, A. P.; Gouveia, L.; et al. Evaluation of Marine Microalga *Diatronema*
1039 *Vlkianum* Biomass Fatty Acid Assimilation in Wistar Rats. Molecules. 2017, 22, 1097.
1040 DOI:10.3390/molecules22071097.
- 1041 [164] Arterburn, L. M.; Oken, H. A.; Hoffman, J. P.; Bailey-Hall, E.; Chung, G.; Rom, D.;
1042 Hamersley, J.; McCarthy, D. Bioequivalence of Docosahexaenoic Acid from Different Algal Oils in

1043 Capsules and in a DHA-fortified Food. *Lipids*. 2007, 42, 1011–1024. DOI: 10.1007/s11745-007-
1044 3019-7.

1045 [165] Hawthorne, K. M.; Abrams, S. A.; Heird, W. C. Docosahexaenoic Acid (DHA)
1046 Supplementation of Orange Juice Increases Plasma Phospholipid DHA Content of Children. *J. Am.*
1047 *Diet. Assoc.* 2009, 109, 708–712. DOI: 10.1016/j.jada.2008.12.024.

1048 [166] Lane, K. E.; Li, W.; Smith, C.; Derbyshire, E. The Bioavailability of an Omega-3-Rich Algal
1049 Oil Is Improved by Nanoemulsion Technology Using Yogurt as a Food Vehicle. In. *J. Food Sci.*
1050 *Technol.* 2014, 49, 1264–1271. DOI: 10.1111/ijfs.12455.

1051 [167] Skrovankova, S. Seaweed Vitamins as Nutraceuticals. In *Marine Medicinal Foods:*
1052 *Implications and Applications, Macro and Microalgae*; Kim, S.K., Ed.; Elsevier: San Diego, 2011;
1053 pp 357–369.

1054 [168] Goh, L. P.; Loh, S. P.; Fatimah, M. Y.; Perumal, K. Bioaccessibility of Carotenoids and T
1055 Tocopherols in Marine Microalgae, *Nannochloropsis* Sp. And *Chaetoceros* Sp. *Mal. J. Nutr.* 2009,
1056 15, 77–86.

1057 [169] Failla, M. L.; Huo, T.; Thakkar, S. K. In Vitro Screening of Relative Bioaccessibility of
1058 Carotenoids from Foods. *Asia. Pac. J. Clin. Nutr.* 2008, 17, 200–203.

1059 [170] Gille, A.; Trautmann, A.; Posten, C.; Briviba, K. Bioaccessibility of Carotenoids from
1060 *Chlorella Vulgaris* and *Chlamydomonas Reinhardtii*. *Int. J. Food Sci. Nutr.* 2016, 67, 507–513.
1061 DOI:10.1080/09637486.2016.1181158.

1062 [171] Tang, G.; Suter, P. M. Vitamin A, Nutrition, and Health Values of Algae: *Spirulina*,
1063 *Chlorella*, and *Dunaliella*. *J. Pharm. Nutr. Sci.* 2011, 1, 111–118. DOI: 10.6000/1927-
1064 5951.2011.01.02.04.

1065 [172] Mendes-Pinto, M. M.; Raposo, M. F. J.; Bowen, J.; Young, A. J.; Morais, R. Evaluation of
1066 Different Cell Disruption Processes on Encysted Cells of *Haematococcus Pluvialis*: Effects on

1067 Astaxanthin Recovery and Implications for Bio-Availability. *J. Appl. Phycol.* 2001, 13, 19–24.
1068 DOI: 10.1023/A:1008183429747.

1069 [173] Okada, Y.; Ishikura, M.; Maoka, T. Bioavailability of Astaxanthin in *Haematococcus* Algal
1070 Extract: The Effects of Timing of Diet and Smoking Habits. *Biosci. Biotechnol. Biochem.* 2009, 73,
1071 1928–1932. DOI: 10.1271/bbb.90078.

1072 [174] Carlsson, A. S.; van Beilen, J. B.; Möller, R.; Clayton, D. EPOBIO Project Report: Micro-
1073 and Macro-Algae: Utility for Industrial Applications; Ed, Bowles, D. CPL Press: UK, 2007 1–86.

1074 [175] Norsker, N.-H.; Barbosa, M. J.; Vermuë, M. H.; Wijffels, R. H. Microalgal Production - A
1075 Close Look at the Economics. *Biotechnol. Adv.* 2011, 29, 24–27. DOI:
1076 10.1016/j.biotechadv.2010.08.005.

1077 [176] Acién, F. G.; Fernández, J. M.; Magán, J. J.; Molina, E. Production Cost of a Real Microalgae
1078 Production Plant and Strategies to Reduce It. *Biotechnol. Adv.* 2012, 30, 1344–1353.
1079 DOI:10.1016/j.biotechadv.2012.02.005.

1080 [177] Ruiz, J.; Olivieri, G.; de Vree, J.; Bosma, R.; Willems, P.; Reith, J. H.; Eppink, M. H. M.;
1081 Kleinegris, D. M. M.; Wijffels, R. H.; Barbosa, M. J. Towards Industrial Products from Microalgae.
1082 *Energy Environ. Sci.* 2016, 9, 3036–3043. DOI: 10.1039/C6EE01493C.

1083 [178] García, J. L.; de Vincente, M.; Galán, B. Microalgae, Old Sustainable Food and Fashion
1084 Nutraceuticals. *Microb. Biotechnol.* 2017, 10, 1017–1024. DOI: 10.1111/1751-7915.12745.

1085 [179] Glemser, M.; Heining, M.; Schmidt, J.; Becker, A.; Garbe, D.; Buchholz, R.; Brück, T.
1086 Application of Light-Emitting Diodes (Leds) in Cultivation of Phototrophic Microalgae: Current
1087 State and Perspectives. *Appl. Microbiol. Biotechnol.* 2016, 100, 1077–1088. DOI:10.1007/s00253-
1088 015-7144-6.

- 1089 [180] Sun, Y.; Huang, Y.; Liao, Q.; Fu, Q.; Zhu, X. Enhancement of Microalgae Production by
1090 Embedding Hollow Light Guides to a Flat-Plate Photobioreactor. *Bioresour. Technol.* 2016, 207,
1091 31–38. DOI: 10.1016/j.biortech.2016.01.136.
- 1092 [181] Zerrouh, O.; Reinoso-Moreno, J. V.; López-Rosales, L.; Cerón-García, M. D. C.; Sánchez-
1093 Mirón, A.; García-Camacho, F.; Sánchez-Mirón, A.; García-Camacho, F.; Molina-Grima, E.
1094 Biofouling in Photobioreactors for Marine Microalgae. *Crit. Rev. Biotechnol.* 2017, 20, 1–18.
- 1095 [182] Khan, M. I.; Shin, J. H.; Kim, J. D. The Promising Future of Microalgae: Current Status,
1096 Challenges, and Optimization of a Sustainable and Renewable Industry for Biofuels, Feed, and
1097 Other Products. *Microb. Cell. Fact.* 2018, 17, 36. DOI: 10.1186/s12934-018-0879-x.
- 1098 [183] Havlik, I.; Scheper, T.; Reardon, K. F. Monitoring of Microalgal Processes. *Adv. Biochem.*
1099 *Eng. Biotechnol.* 2016, 153, 89–142.
- 1100 [184] Reijnders, M. J.; van Heck, R. G.; Lam, C. M.; Scaife, M. A.; Dos Santos, V. A.; Smith, A.
1101 G.; Schaap, P. J. Green Genes: Bioinformatics and Systems-Biology Innovations Drive Algal
1102 Biotechnology. *Trends. Biotechnol.* 2014, 32, 617–626. DOI: 10.1016/j.tibtech.2014.10.003.
- 1103 [185] Al-Thawadi, S.; Public Perception of Algal Consumption as an Alternative Food in the
1104 Kingdom of Bahrain. *J. Basic Appl. Sci. Uni. Bahrain.* 2018, 25, 1–12.

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1111 **Figure caption**

1112 Fig. 1. Possible use of microalgae in foods and beverages.

1113 Fig. 2. Chemical structures of main pigments produced by microalgae. In phycocyanin, the
1114 CH=CH₂ group indicated by the asterisk is replaced by CH₃-CH₂.

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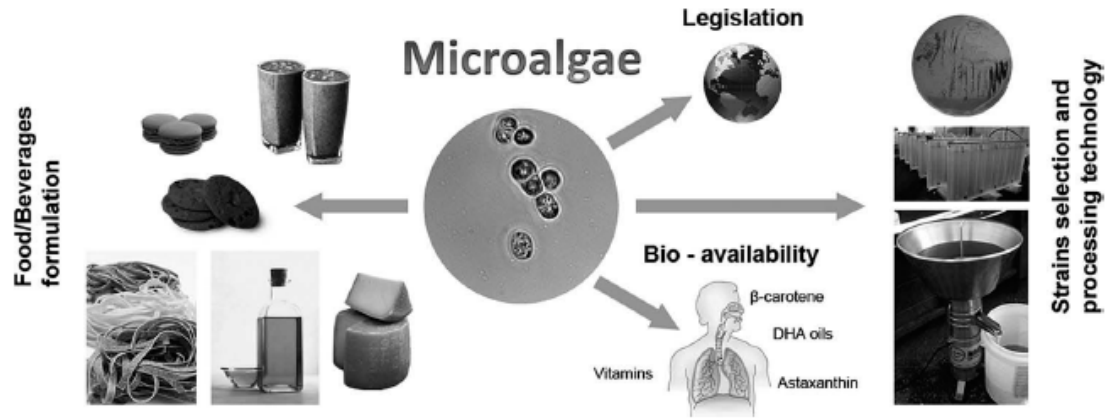
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Fig. 1

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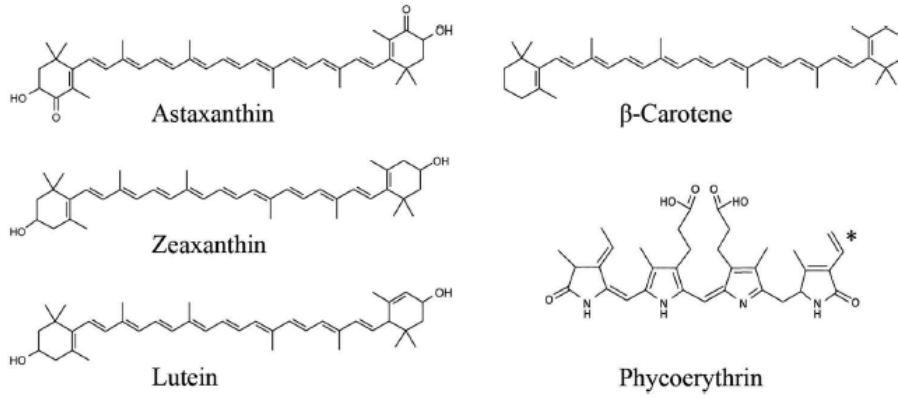
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Fig. 2

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1163 **Table 1.** Chemical profile of different microalgae – proteins, carbohydrates, lipids, essential amino acids, and ω -3 fatty acids.

Microalga	Proteins (% dry matter) ^a	Carbohydrates (% dry matter) ^a	Lipids (% dry matter) ^a	Essential amino acids (g/100 g protein) ^b									ω -3 fatty acids (mg/g dry weight) ^c	
				Ile	Leu	Val	Lys	Met	Cys	Trp	Thr	His	EPA	DHA
<i>Chlorella vulgaris</i> ^a / <i>Chlorella</i> sp. ^{b/c}	51-58	12-17	14-22	4.4	9.2	6.1	8.9	2.2	0.4	-	4.7	2.4	0.4*	-
<i>Dunaliella salina</i>	57	32	6	4.2	11.0	5.8	7.0	2.3	1.2	0.7	5.4	1.8	-	-
<i>Scenedesmus obliquus</i> ^{a,b} / <i>Scenedesmus</i> sp. ^c	50-56	10-17	12-14	3.6	7.3	6.0	5.6	1.5	0.6	0.3	5.1	2.1	0.2*	-
<i>Arthrospira maxima</i>	60-71	13-16	6-7	6.7	9.8	7.1	4.8	2.5	0.9	0.3	6.2	2.2	-	2.2*
<i>Nannochloropsis</i> sp.	47.7	11.2	32.2	4.7	9.4	6.0	6.8	8.4	0.1	-	4.9	2.6	36.8	-
<i>Nitzschia laevis</i> ^c / <i>Nitzschia closterium</i> ^{a,b}	36.2	21.3	35.4	4.8	7.2	5.1	5.6	2.2	0.6	1.7	4.9	2.4	20-40	-
<i>Isochrysis galbana</i>	40.3	19.6	31.1	5.1	9.2	6.1	3.1	2.5	1.0	2.5	4.6	1.7	0.8	15.5
<i>Pavlova</i> sp. ^{a,c} / <i>Pavlova lutheri</i> ^b	24-29	6-9	9-14	4.8	7.7	5.8	6.2	3.1	0.81	3.0	4.5	2.5	18.0	13.2
<i>Phaeodactylum tricornutum</i>	30	8.4	14	4.6	7.0	5.1	6.4	2.7	-	2.6	4.8	1.5	28.4	0.2
<i>Tetraselmis</i> sp. ^{a,c} / <i>Tetraselmis chuii</i> ^b	52	15	16-45	3.4	7.3	4.8	5.6	2.4	-	2.3	4.0	1.6	4.8	0.2
<i>Ulkenia</i> sp.	na	na	na	na	na	na	na	na	na	na	na	na	-	50
<i>Schizochytrium</i> sp.	12.5	38.9	38.9	0.2 [†]	0.8 [†]	0.3 [†]	0.4 [†]	< 0.1 [†]	< 0.1 [†]	< 0.1 [†]	0.5 [†]	0.1 [†]	-	11-24
<i>Cryptocodinium cohnii</i>	12-15	40-50	12-15	na	na	na	na	na	na	na	na	na	-	20-60

^aReferences 20, 53–57

^bReferneces 58–60

^cReferences 59, 65–69

*Expressed as % total fatty acids

[†]Expressed as % dry weight

na, not available

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1167 **Table 2.** Chemical profile of different microalgae – pigments.

Microalga	Pigments (% dry weight)					
	β -carotene	Astaxanthin	Zeaxanthin	Lutein	Allophycocyanin/ C-phycocyanin	B-phycoerythrin /R-phycocyanin
<i>Dunaliella salina</i>	2.7-12.0	0.2	1.1	0.7	np	np
<i>Haematococcus pluvialis</i>	0.04-0.1	4.0-10.0	na	0.8	np	np
<i>Chlorella zofingiensis</i>	na	1.5->4.0	na	0.4	np	np
<i>Auxenochlorella prototheicoides</i>	na	0.08	na	4.6	np	np
<i>Arthrospira platensis</i>	5×10^{-3}	7×10^{-4}	1.3×10^{-4}	6×10^{-6}	9.5/9.6	na
<i>Porphyridium cruentum</i>	na	na	na	na	na	32.7/11.9

References 13,70

na, not available

np, not present

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1176 **Table 3.** Chemical profile of different microalgae – vitamins.

Microalga	Vitamins (mg/100 g dry weight)										
	A	B1	B2	B3	B5	B6	B9	B12	C	E	H
<i>Chlorella</i>	30.8	1.7	4.3	23.8	1.1	1.4	0.094	0.0001	10.4	1.5	-
<i>Isochrysis galbana</i>	127 500*	1.4	3.0	7.8	0.1	0.2	0.3	0.1	11.9	5.8	0.01
<i>Schizochytrium</i> sp.	0.0336	4.4	2.9	14.0	3.5	1.4	0.1	0.0549	-	0.00045	0.3
<i>Arthrospira</i>	0.34	2.4	3.7	12.8	-	0.4	0.094	-	10.1	5.0	-
<i>Tetraselmis suecica</i>	493 750*	3.2	1.9	8.9	3.8	0.3	0.3	0.1	19.1	42.2	0.1
<i>Chlorella stigmatophora</i>	82 300*	1.5	2.0	8.3	2.1	0.2	0.3	0.1	10.0	66.9	0.1
<i>Dunaliella tertiolecta</i>	137 500*	2.9	3.1	7.9	1.3	0.2	0.5	0.1	16.3	11.6	0.1

References 77–79

*Expressed as IU/Kg dry weight (1 IU= 0.6 µg of β-carotene)

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1185 **Table 4.** Microalgae and compounds currently approved as either food or food additives in various countries.

Species/Product	European Union	United States	Australia and New Zealand	Japan
<i>Tetraselmis chuii</i>	Regulation (EC) N° 258/97 (80)			
<i>Ulkenia</i> sp.	Decision 2009/777/EC (85)			
<i>Arthrospira</i> sp. (formerly <i>Spirulina</i>) Phycocyanin from <i>Arthrospira</i> sp. (formerly <i>Spirulina</i>)		GRAS ¹ Notice N° 000101 (2002) (87)		Specifications and Standards for Food Additives (95)
DHA rich oil from <i>Schizochytrium</i> sp.	Commission Implementing Decision (EU) 2015/545 (83) Commission Implementing Decision (EU) 2015/546 (83) Commission Implementing Decision 2014/463/EU (84)	GRAS Notice N° 000137 (2003) (88)	Schedule 25 (2017) (94)	
DHA rich oil from <i>Ulkenia</i> sp.		GRAS Notice N° 000160 (2004) (89)	Schedule 25 (2017) (94)	
<i>Dunaliella salina</i>		GRAS Notice N° 000330 (2010) (90)		
<i>Auxenochlorella protothecoides</i>		GRAS Notice N° 000351 (2010) (91)		
<i>Chlorella vulgaris</i> <i>Chlorella</i> sp.		GRAS Notice N° 000396 (2011) (92)		Specifications and Standards for Food Additives (95)
<i>Euglena gracilis</i> Carotene from <i>Dunaliella</i> sp.		GRAS Notice N° 697 (2017) (93)		Specifications

Haematococcus algae colour

and Standards
for Food
Additives (95)

1186 ¹Generally Recognised As Safe

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1201 **Table 5.** Examples of microalgal food and beverages present on the worldwide market.

Microalgae/ Micro-algal compounds	Genus/Specie	Product/ percentage of incorporation	Company	Health aspects	Technological aspects	References
Dried biomass - AlgaVia® Lipid-Rich Whole Algae	<i>Auxenochlorella protothecoides</i>	Sauce (3%) Dressing (1%) Bakery (4%)	Corbion (The Netherlands)	Reduced saturated fat, rich in fibre, proteins, micronutrients, monounsaturated fatty acids	Enhance taste and texture, emulsification, water binding	http://algavia.com/wp-content/uploads/2017/09/AlgaVia-Lipid-Rich.pdf
Extracted oil - AlgaWise®	Not available	Food oil Algae butter	Corbion (The Netherlands)	Monounstaurated fats and fatty acids (93%),saturated fat (4%), ω-9s Reduced saturated fat, no <i>trans</i> fat	High heat cooking Sharp melting properties	http://algawise.com/
Dried biomass	<i>Arthrospira</i> (formerly <i>Spirulina</i>)	Chocolate bars biscuits, pasta (2%)	Microlife Nutrition (It)	Rich in iron, β-carotene, A,B, K, proteins		https://microlife.bio/collections/alimentazione

		Pasta (2-10%)	Nutracentis (It), La finestra sul cielo (It), Cornand (Fr), GlobeXplore (Fr)			http://www.nutracentis.com/ , http://lafinestrasulcielo.it/it/prodotti/pasta-e-condimenti/pasta-speciale/pasta-alla-spirulina/1/ , http://www.cornand.fr/FR/les-pates-haute-couture/pates-laminees.htm , https://www.algues.fr/fr/boutique/tagliatellines-spiruline
Phycocyanin	<i>Phorphyridium aeruginosum</i>	Pepsi [®] , Bacardi Breezer [®] (na)	PepsiCo. (USA), Bacardi & Co Ltd (Bermuda)		Colour stability	(45)
	<i>Arthrospira</i> (formerly <i>Spirulina</i>)	Linablue [®] (0.01-0.1%)	DIC Corporation (Japan)		Colour/antioxidant	http://www.dic-global.com/us/en/products/health_foods/
EPA/DHA	<i>Chrypthecodinium</i> <i>Nannochloropsis</i> <i>Odontella</i> <i>Schizochytrium</i>	Food ingredient	Martek/DSM (USA/NL) Blue Biotech (Germany) InnovaIG (Fr) Xiamen Huison Biotech Co. (China) Cellana (Hawaii)	Anti-inflammatory, Protection against cardiovascular disease, atherosclerosis	Avoiding fish oil taste	(3)
	Not available					http://cellana.com

β -carotene	<i>Dunaliella</i>	Food additive, colourant	Cognis Australia BASF (Au/DE) Tianjin Lantai Laboratory (China) Nature Beta Technologies (Israel) Pro Algen (India)	Anti- inflammatory Antioxidant	Natural colour	(3)
Astaxanthin	<i>Haematococcus</i>	Food ingredient, additive	Algatech (India) Blue Biotech (Germany) Fuji Chemicals (Japan) Mera Pharma (USA) BioReal (Sweden)	Anti- inflammatory Antioxidant		(3)